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(54) **METHOD OF USING ATTENUATED  
BORDETELLA STRAINS**

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30, 2006, provisional application No. 60/780,827,  
filed on Mar. 10, 2006.

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*A61K 39/02* (2006.01)  
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CPC ..... *A61K 39/099* (2013.01); *A61K 2039/522*  
(2013.01); *A61K 2039/523* (2013.01); *A61K*  
*2039/543* (2013.01)

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None  
See application file for complete search history.

(56) **References Cited**

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(57) **ABSTRACT**

A mutated *Bordetella* strain comprising at least a mutated ptx  
gene, a deleted or mutated dnt gene and a heterologous ampG  
gene is provided. The attenuated mutated *Bordetella* strain  
can be used in an immunogenic composition or a vaccine for  
the treatment or prevention of a *Bordetella* infection. Use of  
the attenuated *Bordetella* strain for the manufacture of a vac-  
cine or immunogenic composition, as well as methods for  
protecting mammals against infection by *Bordetella* are also  
provided.

**7 Claims, 14 Drawing Sheets**

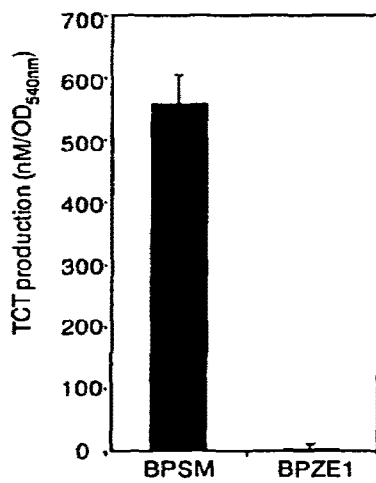


Fig. 1

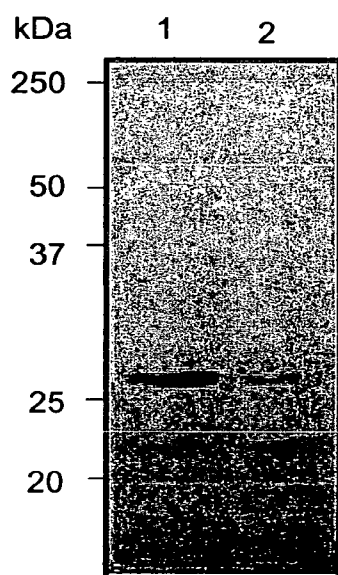


Fig. 2

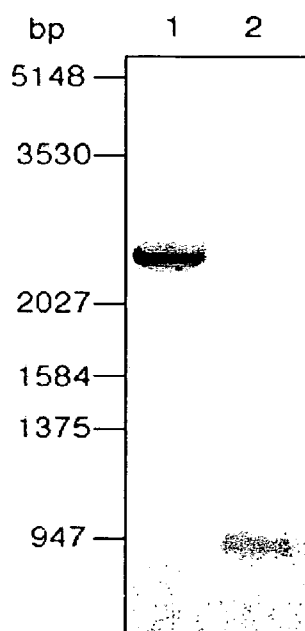


Fig. 3

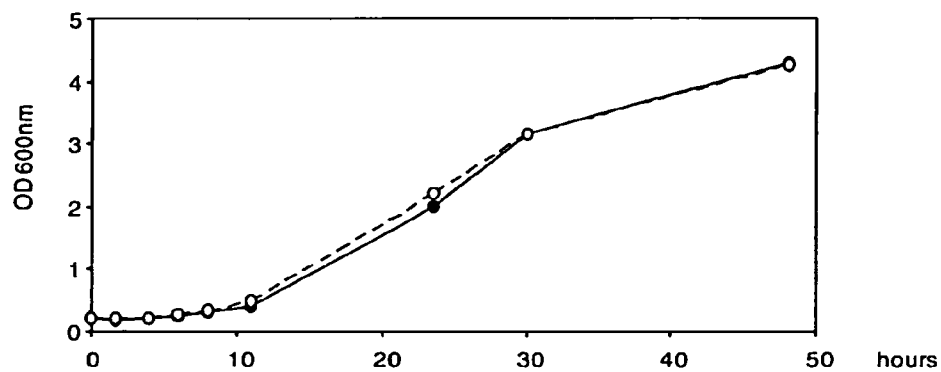


Fig. 4

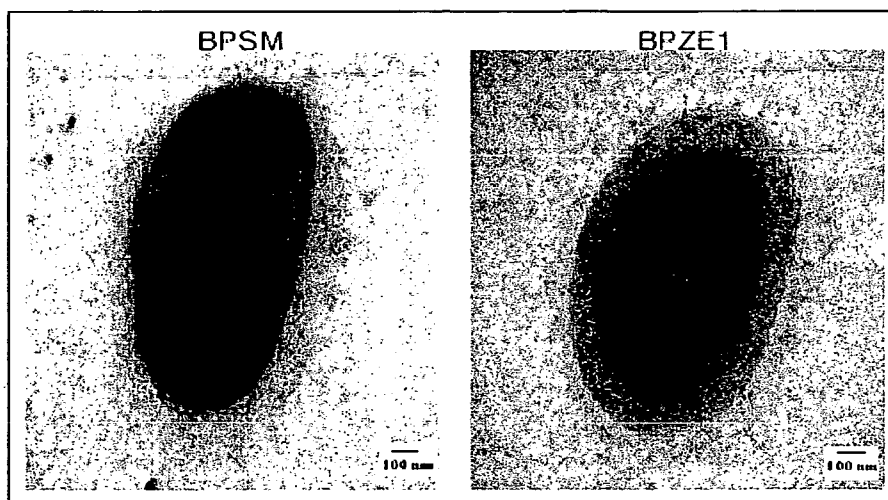


Fig. 5

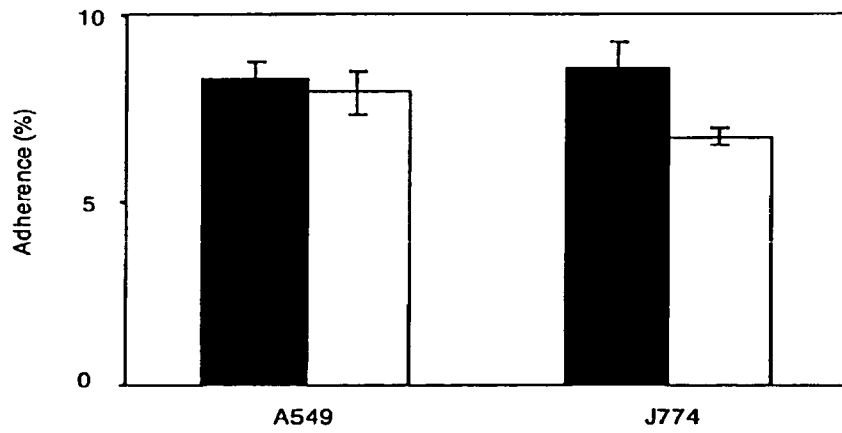


Fig. 6

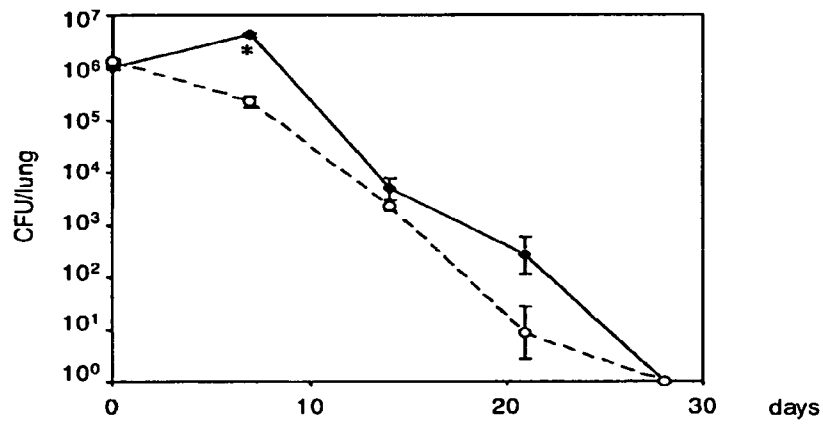


Fig. 7



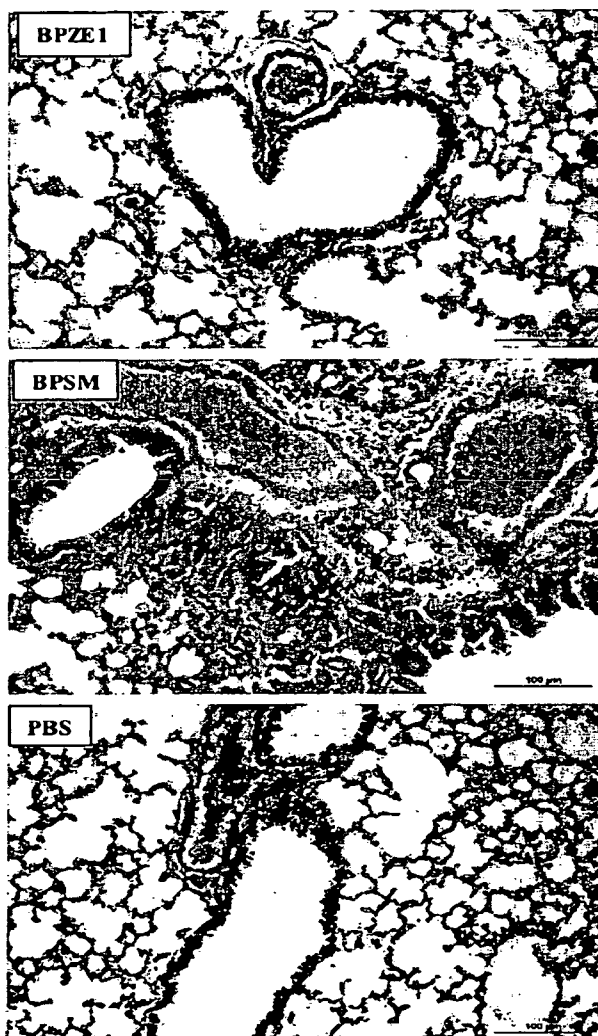


Fig. 8

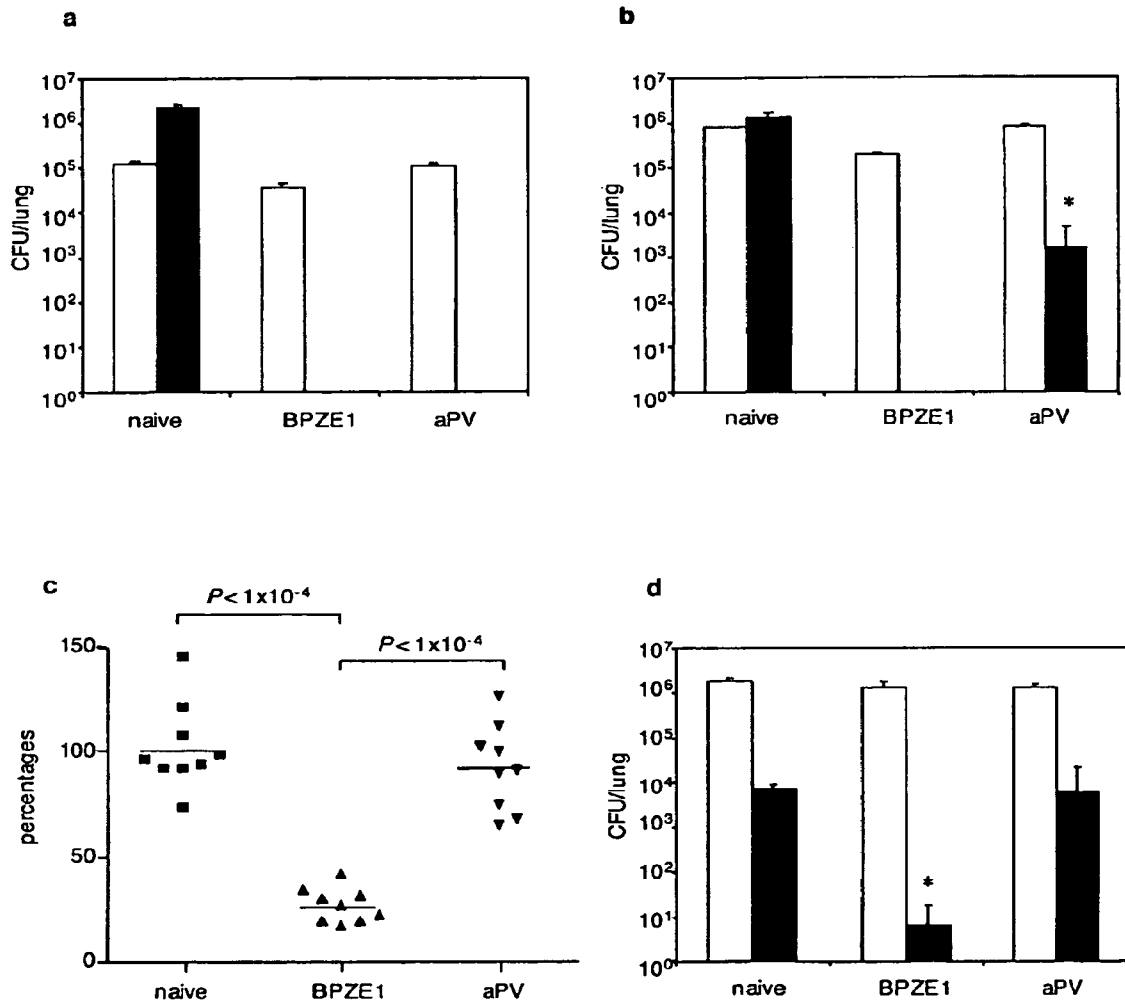


Fig. 9

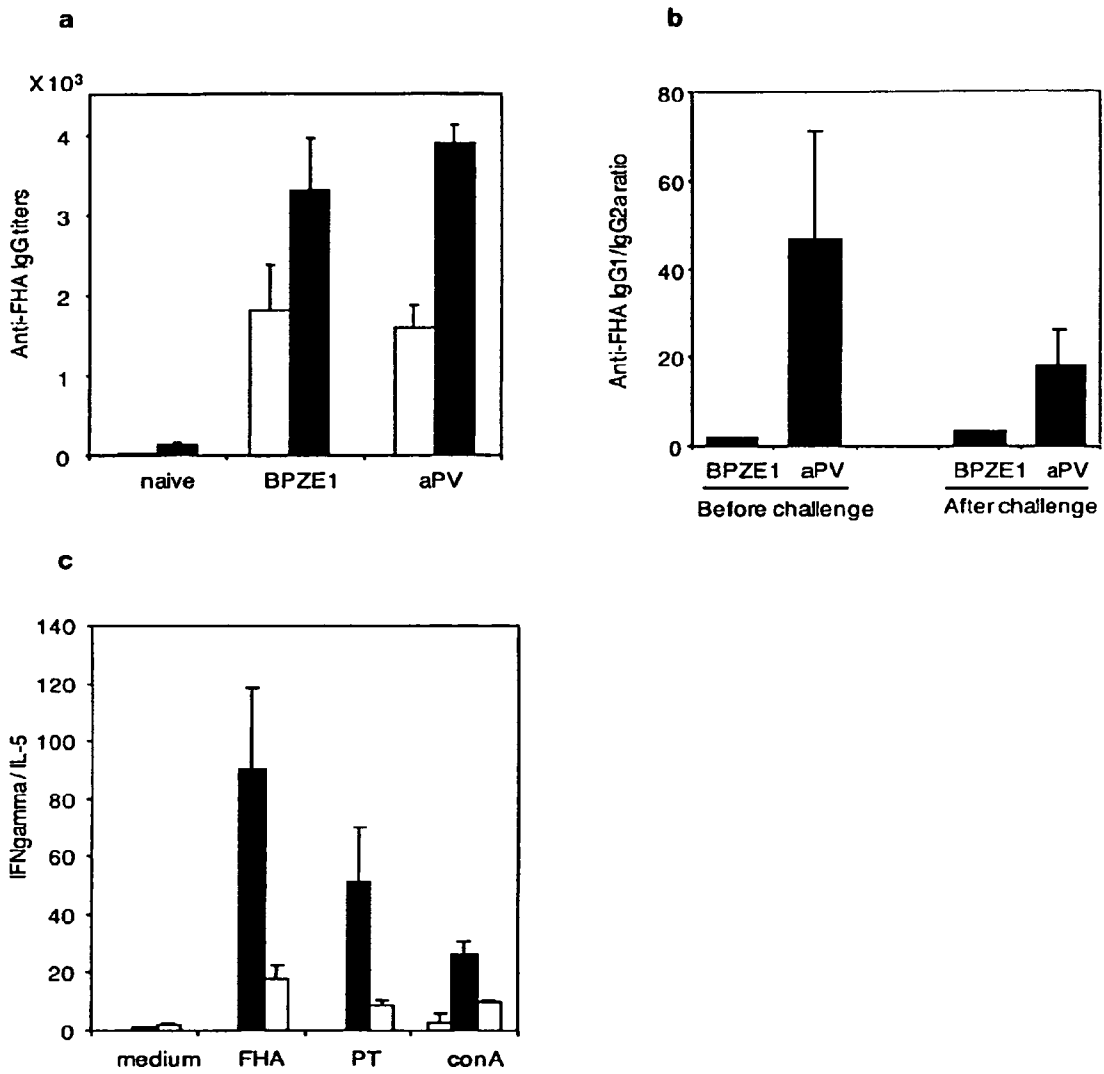


Fig. 10

**Islet-activating protein S1 (NP\_882282)**

MRCTRAIRQTARTGWLTLA I LAVTAPVTSPAWADDPPATVYRYDSRPPEDVF  
QNGFTAWGNNDNVLDHLTGRSCQVGSSNSAFVSTSSSRRYTEVYLEHRMQEAV  
EAERAGRGTGHFIFYIYEVRADNNFYGAASSYFEYVDTYGDNAGRILAGALAT  
YQSEYLAHRRIPPENIRRVTRVYHNGITGETTTTEYSNARYVSQQTRANPNPY  
TSRRSVASIVGTLVRMAPVIGACMARQAESSEAMAASERAGEAMVLVYYESI  
AYSF

Fig. 11

**Dermonecrotic toxin (NP\_881965)**

MDKDESALRQLVDMALVGYDGVVEELLALPSEESGDLAGGRAKREKAEFALFS  
EAPNGDEPIGQDARTWFYFPHYRPAVAVSNLKKMQVAIRARLEPESLILQWLIA  
LDVYLGVLIAALSRTVISDLVFEYVKARYEIIYLLNRVPHPLATAYLKRRRQR  
PVDRSGRLGSVFEHPLWFAYDELAGTVDLADADIYEQALAESIERMDGEPDDG  
SLDTAEHDVWRLCRDGINRGEQAIFQASGPYGVVADAGYMRTVADLAYADALA  
DCLHAQLRIRAQGSVDS PGDEMPRKLDaweIAKFHLAATQQARVDLLEAAFAL  
DYAALRDVRVYGDYRNALALRFIKREALRLLGARRGNASTMPAVAAGEYDEIV  
ASGAANDAAYVSMAAALIAGVLCDESARTLPVVLARFRPLGVLARFRRLEQ  
ETAGMLLDGQEPERGFISFTDFRSDAFASYAEYAAQFNIDYIDQYSILEAQR  
LARILALGSRMTVDQWCLPLQKVRHYKVLTSQPGLIARGIENHNRGIEYCLGR  
PPLTDLPLGLFTMFQLHDSSWLLVSNINGELWSDVLANAEMQNPTLAALAEPO  
GRFRTGRRRTGGWFLGGPATEGPSLRDNYLLKLRQSNPGLDVKKCWYFGYRQY  
RLPAGALGVPLFAVSVALRHSLLDAAHAKSALYKPSEWQKFAFWIVPFYREI  
FFSTQDRSYRVDVGSIVFDSISLLASVFSIGGKLSFTRTQYGNLRNFVVRQR  
IAGLSGQRLWRSVLKELPALIGASGLRSLRSLLDVLYEIFEPVPIRRLVAGFV  
SATTVGGRNQAFRLRQAFSAASSAGRTGGQLASEWRMAGVDATGLVESTSGGR  
FEGIYTRGLGPLSECTEHFIVESGNAYRVIWDAYTHGWRVNVNRLPPRLTYTV  
PVRLNGQGHWETHLDVPGRGGAPEIFGRIRTRNLVALAAEQAAPMRLLNQAR  
RVALRHIDTCRSRLALPRAESDMDAAIRIFFGEPDAGLRQRIGRRLQEVRAI  
GDLSPVNDVLYRAGYDLDDVATLFFNAVDRNTSLGRQARMELYLDAIVDLHARL  
GYENARFVDLMAFHLLSLGHAATASEVVEAVSPRLLGNVFDISNVAQLERGIG  
NPASTGLFVMLGAYSESSPAIFQSFVNDIFPAWRQASGGGPLVWNFGPAAISP  
TRLDYANTDIGLLNHGDISPLRARPPGGRRDIDLPPGLDISFVRYDRPVRMS  
APRALDASVFRPVDGPVHGYIQSWTGAEIEYAYGAPAAAREVMLTDNVRISI  
ENGDEGAIGVRVRLDTPVATPLILTGGSLSGCTTMVGVKEGYLAFYHTGKST  
ELGDWATAREGVQALYQAHLAMGYAPISIPAPMRNDDLVSIAATYDRAVIAYL  
GKDVPGGGSTRITRHDEGAGSVVSFDYNAAVQASAVPRLGQVYVLI SNDGQGA  
RAVLLAEDLAWAGSGSALDVLNERLVTLFPAPV

Fig. 12

**AmpG protein (NP\_878961.1)**

MAPLLVLGFASGLPLALSSGTLQAWATVENVSLQSIGFLTLAGTAYTLKFLWA  
PLIDRYVPPFLGRRRGWMLLTQVLLAAAIMVMGMLSPGSALLPLALVAVLVAF  
LSASQDIAFDAYSTDVLRQEERGAGAAAMRVMGYRLAMIVSGGLALIVADRWL  
WGNTYVLMGGLMLACALGTLWAPERPERPANPPRDLGAAVVEPFREFFSRRGAI  
DMLLLIVLYKLGDAFAGALSTTFLLRGAGFSATEVGTVNKVLGLAATIVGALA  
GGSIMTRWGLYRSLMAFGLLQAVSNLGYWLIAVSPKNLYLMGLAVGVENLCGG  
LGTASFVALLMAMCRQQFSATQFALLSALAAVGRTYLAGPLTPVLVEWLDWPG  
FFIVTVLIALPGLWLLRLRRNVIDELDAQAR

Fig. 13

**AmpG protein (NP\_752478.1)**

MSSQYLRI FQQPRSA ILLI LGFASGLPLALTSGLTQAWMTVENIDLKTIGFFS  
LVGQAYVFKFLWSPLMDRYTPPFFGRRRGWLLATQIILLV AIAAMGFLEPGTQ  
LRWMAALAVVIAFCSASQDIVFDAWKTDVLP AEERGAGAAISVLGYRLGMLVS  
GGLALWLADKWLGWQGM YWLMAALLIPCI IATLLAPEPTDTIPVPKTLEQAVV  
APLRDFFGRNNAWL ILLLI VLYKLGDAFAMSLTTTFLIRGVGFDAGEVGVV NK  
TLGLLATIVGALYGGILMQRLSLFRALLIFGILQGASNAGYWLLSITDKHLYS  
MGAAVFFENLCGGMGTS AFVALLMTLCNKSF SATQFALLSALS AVGRVYVGPV  
AGWFVEAHGWSTFYLF SVAAAVPGL ILLLVCRQTLEYTRVNDNFISRTEYPAG  
YAFAMWTLAAGISLLAVWLLLLTMDALDLTHFSFLPALLEVGV LVALSGVVLG  
GLLDYLALRKTHLM

Fig. 14

## METHOD OF USING ATTENUATED BORDETELLA STRAINS

### CROSS-REFERENCE TO RELATED APPLICATIONS

The present application is a divisional application of U.S. nonprovisional patent application Ser. No. 12/224,895 filed on Nov. 19, 2008 as a national stage entry application under 35 U.S.C. 371 of international patent application number PCT/EP/001942, filed on Mar. 7, 2007, which designated the U.S. and claims the priority of U.S. provisional patent application Ser. No. 60/817,430 filed on Jun. 30, 2006 and U.S. provisional patent application Ser. No. 60/780,827 filed on Mar. 10, 2006, which are incorporated herein by reference in their entirety.

### FIELD OF THE INVENTION

The present invention relates to a mutated *Bordetella* strain comprising at least a mutated *ptx* gene, a deleted or mutated-*dnt* gene and a heterologous *ampG* gene. The attenuated mutated *Bordetella* strain can be used in an immunogenic composition or a vaccine for the treatment or prevention of a *Bordetella* infection. Use of the attenuated *Bordetella* strain for the manufacture of a vaccine or immunogenic compositions, as well as methods for protecting mammals against infection by *Bordetella* also form a part of the invention.

### BACKGROUND OF THE INVENTION AND RELATED PRIOR ART

*Pertussis* is still among the principal causes of death worldwide, and its incidence is increasing even in countries with high vaccine coverage. Although all age groups are susceptible, it is most severe in infants too young to be protected by currently available vaccines.

Whooping cough or *pertussis* is a severe childhood disease responsible for high mortality rates before the introduction of effective vaccines in the second half of the 20th century. The success of these vaccines has led to the opinion that the disease is essentially under control, although world-wide 200,000 to 400,000 *pertussis*-linked deaths are still recorded annually, and the disease still ranks sixth among the causes of mortality due to infectious agents [1]. Although mostly prevalent in developing countries, the disease is also re-emerging in the developed world [2, 3], including the U.S.A., where the incidence has increased five-fold over the last twenty years [4]. Unexpectedly, the epidemiology of *pertussis* has changed in countries with high vaccine coverage, where cases of adolescent and adult *pertussis* are increasingly frequent [5]. This is probably due to progressive waning of vaccine-mediated immunity during adolescence. Often atypical and therefore difficult to diagnose, *pertussis* is generally not life-threatening in adults and in many cases remains unnoticed. However, infected adults constitute an important reservoir for transmission of the disease to very young children, too young to be fully vaccinated, and therefore at risk to develop severe disease associated with high mortality rates.

*Pertussis* vaccination usually begins at two months of age, and full protection requires at least three immunizations at one- to two-month intervals. Therefore, infants are not fully protected before the age of 6 months using the currently available vaccines. To reduce the incidence of *pertussis* in the very young and most vulnerable age groups, early immunization, possibly at birth, would thus be highly desirable. However, numerous studies in humans and in animal models

have suggested that the neonatal immune system is too immature to effectively induce vaccine-mediated protective immunity [6, 7]. Especially the IFN- $\gamma$  production, indicative of a Th1 response that is essential to the development of protective immunity to *pertussis* [8], appears to be significantly reduced in human newborns, compared to older children or adults [9]. This is also reflected by the fact that significant amounts of antigen-specific IFN- $\gamma$  are only produced after several months ( $\geq 6$  months) in children vaccinated with *pertussis* vaccines, especially with acellular vaccines (aPV) [10].

Natural infection with *Bordetella pertussis* has long been considered to induce strong and long-lasting immunity, that wanes much later than vaccine-induced immunity [5, 11]. Furthermore, infection with *B. pertussis* induces measurable antigen-specific Th1 type immune responses even in very young children (as young as one month of age) [12]. These observations suggest that live vaccines applicable by the nasal route in order to mimic as closely as possible natural infection, may be attractive alternatives over the currently available vaccines.

There are many vaccinating compositions to treat *Bordetella* infections known in the art. However, these immunogenic compositions are not used to treat newborn children or in cases where an epidemic and rapid protective immunity is required.

Thus, French Patent FR 0206666 discloses live *Bordetella* strains that have been rendered deficient in at least two toxins chosen from PTX, DNT, AC and TCT. This patent discloses the over expression of an endogenous *ampG* gene by the addition of a strong promoter, and the addition of 11 terminal amino acids of the *ampG* gene from *E. coli*.

Mielcarek et al, Vaccine (2006; 24(52): 52154-52-55) disclose a strain of *Bordetella pertussis* attenuated of PTK, DTN- and TCr for use in the immunization of mice. This reference discloses that to reduce the production of tracheal cytotoxin, the *ampG* gene should be overexpressed. However, upon further evaluation, the authors realized that by overexpressing the *ampG* gene, there is an increase in tracheal cytotoxin and not a decrease as was originally thought.

Mielcarek et al in Advance Drug Delivery Review 51 (2001) pgs. 55-69 disclose that live vaccines can induce systemic and mucosal responses when administered by the oral or nasal route.

Roduit et al in Infection and Immunity (2002 July; 70(7): 3521-8) describe vaccinating neonatals and infants with mutated *Bordetella* strains with a DTP composition.

Mattoo et al, in Frontiers of Bioscience 6, e168-e186 (2001), suggest replacing the endogenous *ampG* gene in *Bordetella* with the *E. coli ampG* gene, which resulted in a decrease in the amount of TCT produced.

Thus, the prior art although disclosing various types of vaccinating compositions fails to address the problem of providing a vaccine or immunogenic composition that can provide protection to a newborn prior to six months. Furthermore, the prior art fails to disclose an immunogenic or a vaccine that provides rapid protective immunity against a *Bordetella* infection. The prior art also fails to disclose an immunogenic composition or vaccine that provides a rapid protective immunity against a *Bordetella* infection, said protective immunity increasing over at least the next two months following vaccination.

Therefore, it is an object of the present invention to overcome the deficiencies in the prior art.

It is another object of the present invention to produce a live attenuated vaccine candidate or immunogenic composition through genetic attenuation of a *Bordetella* strain such as *B.*



*pertussis* or *B. parapertussis* to diminish pathogenicity, while maintaining the ability to colonize and induce protective immunity.

It is another object of the present invention to produce a vaccine or immunogenic composition that induces protection in newborns after a single intranasal administration that is superior to the protection provided by the current aPV.

It is yet another object of the present invention to provide protection against infection with *Bordetella parapertussis*, as well as *Bordetella pertussis* which was not seen after vaccination with aPV.

Another object of the present invention is to induce strong protective immunity in newborns against *Bordetella* infection.

Yet another object of the present invention is to provide a vaccine or immunogenic composition that induces mucosal and systemic immunity.

It is another object of the present invention to produce a live attenuated *Bordetella pertussis* strain to be given as a single-dose nasal vaccine in early life, called BPZE1.

It is yet another object of the present invention to provide a vaccine that can not only be used to vaccinate newborns, but can be used in all mammals of any age in the case of an epidemic of whooping cough.

Another object of the present invention is to provide a vaccine against *Bordetella* infection that induces a rapid protective immunity and/or a protective immunity that increases over at least the next two months after the vaccination.

Yet another object of the present invention is to provide prevention or treatment against *Bordetella* infection that is relatively low in production costs.

These and other objects are achieved by the present invention as evidenced by the summary of the invention, description of the preferred embodiments and the claims.

### SUMMARY OF THE INVENTION

The present invention provides a mutated *Bordetella* strain comprising at least a mutated *pertussis* toxin (ptx) gene, a deleted or mutated dermonecrotic toxin (dnt) gene, and a heterologous ampG gene.

In another aspect the present invention relates to an immunogenic composition comprising a mutated *Bordetella* strain comprising at least a mutated *pertussis* toxin (ptx) gene, a deleted or mutated *pertussis* dermonecrotic toxin (dnt) gene, and a heterologous ampG gene.

In yet another aspect the present invention provides a vaccine comprising the attenuated *Bordetella* strain comprising at least a mutated *pertussis* toxin (ptx) gene, a deleted or mutated *pertussis* dermonecrotic toxin (dnt) gene, and a heterologous ampG gene.

In still another aspect, the present invention provides the use of an attenuated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene for the manufacture of a vaccine for the prevention of a *Bordetella* infection.

In yet another aspect, the present invention provides the use of an attenuated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene for the manufacture of a vaccine for the induction of an immune response directed preferentially toward the Th1 pathway against said attenuated *Bordetella*.

Also provided is a method of protecting a mammal against disease caused by infection by *Bordetella pertussis* and *Bordetella parapertussis* comprising administering to said mammal in need of such treatment a mutated *Bordetella* strain

comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene.

A method of providing a rapid protective immunity against a *Bordetella* infection comprising administering to said mammal in need of such treatment a mutated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene is also part of the present invention.

A method of providing a rapid protective immunity against a *Bordetella* infection comprising administering to a mammal in need of such treatment a mutated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene or a vaccine comprising said mutated *Bordetella* strain, wherein said method provides further an increase in said protective immunity over at least two months after vaccination is still another aspect of the present invention.

Use of the mutated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene and a heterologous ampG gene for the preparation of a multivalent vaccine (i.e., a vaccine for preventing or treating infections caused by different pathogens) to treat respiratory diseases is yet another aspect of the present invention.

Use of an attenuated *Bordetella* strain of the invention, by administration to mammals in need of a rapid protective immunity against a *Bordetella* infection, wherein said protective immunity increases over at least two months after administration, is also part of the present invention.

A method to provide a mucosal response and a systemic response to treat or protect against *Bordetella* infections in mammals is still another aspect of the present invention.

### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a bar graph illustrating the TCT present in culture supernatants of BPSM and BPZE1 expressed as means of nM/OD<sub>540 nm</sub> ( $\pm$ standard error) of 3 separate cultures for each strain.

FIG. 2 is an immunoblot analysis of PTX production in the culture supernatants of BPSM (lane 1) and BPZE1 (lane 2). The sizes of the Mr markers are expressed in kDa and given in the left margin.

FIG. 3 is a Southern-blot analysis of the dnt locus in BPSM (lane 1) and BPZE (lane 2). The lengths of the size markers are indicated in base pairs (bp) are shown in the left margin.

FIG. 4 is a graph illustrating the growth rates of BPSM (black line) and BPZE1 (dotted line) in liquid culture.

FIG. 5 are electron micrographs representative of BPSM (left) and BPZE1 (right) grown in liquid medium for 24 h.

FIG. 6 is a graph illustrating the in vitro adherence of BPSM (black columns) and BPZE1 (white columns) to human pulmonary epithelial A549 cells (left) and murine macrophage-like J774 cells (right). The results are expressed as means of percentages of binding bacterial relative to the bacteria present in the inoculum from three different experiments.

FIG. 7 is a graph illustrating lung colonization by BPSM (black lines) and BPZE1 (dotted lines) of adult mice infected intranasally with 10<sup>6</sup> CFU of BPZE1 or BPSM. The results are expressed as mean ( $\pm$ standard error) CFUs from three to four mice per group and are representative of two separate experiments. P=0.004.

FIG. 8 are photographs of a histological analysis of lungs from BPZE1 (upper panel) or BPSM-infected (middle panel) adult mice compared to controls given PBS (lower panel). One week after infection, the lungs were aseptically removed

and fixed in formaldehyde. Sections were stained with hematoxylin and eosin and examined by light microscopy.

FIG. 9 are graphs illustrating the protection against *B. pertussis* in (a) adult and (b) infant mice or *B. paraper-tussis* in infant mice (d). Mice immunized with BPZE1, aPV or PBS (naive) were challenged with BPSM (a and b) or *B. paraper-tussis* (d), and lung CFU counts were determined 3 h (white bars) or 7 days (black bars) later. Results are expressed as mean ( $\pm$ standard error) CFUs from 3-4 mice per group and are representative of two separate experiments. (b,\*, P=0.009; d,\*, P=0.007) (c) CFU counts 3 h after BPSM challenge in adult mice vaccinated with BPZE1 or aPV, compared to controls. Results obtained from 3 separate experi-ments are expressed as percentages of CFUs of each mouse relative of the average of CFUs in non-immunized group from the same experiment.

FIG. 10 are bar graphs illustrating the immune responses induced by BPZE1 or aPV immunization. (a) Anti-FHA IgG (H+1) titers and (b) IgG1/IgG2a ratios before (white bars) or 1 week after BPSM challenge (black bars) in BPZE1 or aPV immunized mice, compared to controls. (c) IFN- $\gamma$  to IL-5 ratios produced by FHA-, PTX- or ConA-stimulated spleno-cytes from mice vaccinated 2 months before with BPZE1 (black bars) or aPV (white bars), compared to controls (gray bars). Antibodies and cytokines were measured in individual mice, and the results are expressed as mean values ( $\pm$ standard error) for 4 mice per group tested in triplicate.

FIG. 11 is the amino acid sequence of *pertussis* toxin (SEQ ID NO:1) (islet-activating protein 51). The first 34 amino acids are the signal sequence, while amino acids 35 to 269 are the mature chain.

FIG. 12 is the amino acid sequence of dennonecrotic toxin (SEQ ID NO:2).

FIG. 13 is the amino acid sequence of AmpG from *Bor-detella pertussis* (SEQ ID NO:3).

FIG. 14 is the amino acid sequence of AmpG from *Escheri-chia coli* (SEQ ID NO:4).

#### DESCRIPTION OF THE PREFERRED EMBODIMENTS OF THE PRESENT INVENTION

As used herein, the abbreviation "PTX" refers to *pertussis* toxin, which synthesizes and secretes an ADP-ribosylating toxin. PTX is composed of six polypeptides S1 to S5, the enzymatically active moiety is called S1. PTX has a 34 amino acid signal sequence, while the mature chain consists of amino acids 35 to 269. PTX is the major virulence factor expressed by *B. pertussis*. The A moiety of these toxins exhibit ADP-ribosyltransferase activity and the B portion mediates binding of the toxin to host cell receptors and the translocation of A to its site of action (57).

As used herein the abbreviation "DNT" refers to *pertussis* dermonecrotic toxin, which is a heat labile toxin that induces localized lesions in mice and other laboratory animals when it is injected intradermally. It is lethal to mice when it is injected in low doses intravenously (58 to 61). DNT is considered to be a virulence factor for the production of turbinate atrophy in porcine atrophic rhinitis (62, 63).

As used herein the abbreviation "TCT" refers to tracheal cytotoxin, which is a virulence factor synthesized by *Bor-detellae*. TCT is a peptidoglycan fragment and has the ability to induce interleukin-1 production and nitric oxide synthase. It has the ability to cause stasis of cilia and has lethal effects on respiratory epithelial cells.

The term "mammal" encompasses any of various warm-blooded vertebrate animals of the class Mammalia, including

humans, characterized by a covering of hair on the skin and, in the female, milk-producing mammary glands for nourishing the young.

The term "attenuated" means a weakened, less virulent *Bordetella* strain that is capable of stimulating an immune response and creating protective immunity, but does not cause any illness.

The terminology "rapid protective immunity" means that immunity against *Bordetella* is conferred in a short time after administration of the mutated *Bordetella* strain of the present invention. By "short time" means vaccinated and challenged one week later. More specifically, there is a quick expansion of existing pathogen-specific peripheral lymphocytes, CD4+ cytotoxic effectors (CTLs) and CD4+ helper cells. The CD4+ helper cells induce B cell maturation and antibody production. Thus, lymphocytes with the memory pool are poised to rapidly proliferate at the time of subsequent infection.

The term "*Bordetella* strain" encompasses strains from *Bordetella pertussis*, *Bordetella paraper-tussis* and *Bordetella bron-chiseptica*.

The expression "*Bordetella* infection" means an infection caused by at least one of the three following strains: *Bor-detella pertussis*, *Bordetella paraper-tussis* and *Bordetella bron-chiseptica*.

By "child" is meant a person or a mammal between 6 months and 12 years of age.

By the term "newborn" is meant a person or a mammal that is between 1 day old and 24 weeks of age.

The term "treatment" as used herein is not restricted to curing a disease and removing its causes but particularly covered means to cure, alleviate. Remove or lessen the symp-toms associated with the disease of interest, or prevent or reduce the possibility of contracting any disorder or malfunc-tion of the host body.

The terms "protection" and "prevention" are used herein interchangeably and mean that an infection by *Bordetella* is impeded.

"Prophylaxis vaccine" means that this vaccine prevents *Bordetella* infection upon future exposure.

By "preferentially towards the Th1 pathway" is meant that the Th1 pathway is favored over the Th2 pathway.

The term "immunogenic composition" means that the composition can induce an immune response and is therefore antigenic. By "immune response" means any reaction by the immune system. These reactions include the alteration in the activity of an organism immune system in response to an antigen and may involve, for example, antibody production, induction of cell-mediated immunity, complement activation or development of immunological tolerance

More specifically, the present invention provides at least a triple mutated *Bordetella* strain that can be used as an immu-nogenic composition or a vaccine. It will be appreciated that the at least triple mutated *Bordetella* strain contains a mutated ptx gene, a deleted or mutated dnt gene and a heterologous ampG gene. The heterologous ampG gene product reduces in large quantities the amount of tracheal cytotoxin that is pro-duced.

The present invention is not limited to only the triple mutants described above. Other additional mutations can be undertaken such as adenylate cyclase (AC) deficient mutants (64), lipopolysaccharide (LPS) deficient mutants (65), fila-mentous hemagglutinin (FHA) (66) and any of the bvg-regu-lated components (67).

The starting strain which is mutated can be any *Bordetella* strain including *Bordetella pertussis*, *Bordetella paraper-tus-*

*sis* and *Bordetella bronchiseptica*. In one aspect the starting strain used to obtain the mutated *Bordetella* strain is *B. pertussis*.

The construction of the mutated *Bordetella* strain starts with replacing the *Bordetella* ampG gene in the strain with a heterologous ampG gene. Any heterologous ampG gene can be used in the present invention. These include all those gram-negative bacteria that release very small amounts of peptidoglycan fragments into the medium per generation. Examples of gram-negative bacteria include, but are not limited to *Escherichia coli*, *Salmonella*, *Enterobacteriaceae*, *Pseudomonas*, *Moraxella*, *Helicobacter*, *Stenotrophomonas*, *Legionella* and the like.

By replacing the *Bordetella* ampG gene with a heterologous ampG gene, the amount of tracheal cytotoxin (TCT) produced in the resulting strain expresses less than 1% residual TCT activity. In another embodiment, the amount of TCT toxin expressed by the resulting strain is between 0.6% to 1% residual TCT activity or 0.4% to 3% residual TCT activity or 0.3% to 5% residual TCT activity.

PTX is a major virulence factor responsible for the systemic effects of *B. pertussis* infections, as well as one of the major protective antigens. Due to its properties, the natural ptx gene is replaced by a mutated version so that the enzymatically active moiety S1 codes for an enzymatically inactive toxin, but the immunogenic properties of the *pertussis* toxin are not affected. This can be accomplished by replacing the lysine (Lys) at position 9 of the sequence with an arginine (Arg). Furthermore, a glutamic acid (Glu) at position 129 is replaced with a glycine (Gly).

Other mutations can also be made such as those described in U.S. Pat. No. 6,713,072, incorporated herein by reference, as well as any known or other mutations able to reduce the toxin activity to undetectable levels. Allelic exchange is first used to delete the ptx operon and then to insert the mutated version.

Finally, the dnt gene is then removed from the *Bordetella* strain by using allelic exchange. Besides the total removal, the enzymatic activity can also be inhibited by a point mutation. Since DNT is constituted by a receptor-binding domain in the N-terminal region and a catalytic domain in the C-terminal part, a point mutation in the dnt gene to replace Cys-1305 to Ala-1305 inhibits the enzyme activity of DNT (68). DNT has been identified as an important toxin in *Bordetella bronchiseptica* and displays lethal activity upon injection of minute quantities (26).

Besides allelic exchange to insert the mutated ptx gene and the inhibited or deleted dnt gene, the open reading frame of a gene can be interrupted by insertion of a genetic sequence or plasmid. This method is also contemplated in the present invention.

The triple mutated strain of the present invention is called a BPZE1 strain and has been deposited with the Collection Nationale de Cultures de Microorganismes (CNCM) in Paris, France on Mar. 9, 2006 under the number CNCM 1-3585. The mutations introduced into BPZE1 result in drastic attenuation, but allow the bacteria to colonize and persist. Thus, in another embodiment the present invention provides BPZE1, which can induce mucosal immunity and systemic immunity when administered. In another aspect the BPZE1 is administered intranasally.

The mutated *Bordetella* strains of the present invention can be used in immunogenic compositions. Such immunogenic compositions are useful to raise an immune response, either an antibody response and or preferably a T cell response in

mammals. Advantageously, the T cell response is such that it protects a mammal against *Bordetella* infection or against its consequences.

The mutated *Bordetella* strains of the present invention can be used as live strains or chemically or heat-killed strains in the vaccines or immunogenic compositions. In one aspect, the live strains are used for nasal administration, while the chemically—or heat killed strains can be used for systemic or mucosal administration.

The immunogenic composition may further comprise a pharmaceutically suitable excipient or carrier and/or vehicle, when used for systemic or local administration. The pharmaceutically acceptable vehicles include, but are not limited to, phosphate buffered saline solutions, distilled water, emulsions such as an oil/water emulsions, various types of wetting agents sterile solutions and the like.

The immunogenic composition of the invention can also comprise adjuvants, i.e., any substance or compound capable of promoting or increasing a T-cell mediated response, and particularly a CD4<sup>+</sup>-mediated or CD8<sup>+</sup>-mediated immune response against the active principle of the invention. Adjuvants such as muramyl peptides such as MDP, IL-12, aluminium phosphate, aluminium hydroxide. Alum and/or Montanide® can be used in the immunogenic compositions of the present invention.

It would be appreciated by the one skilled in the art that adjuvants and emulsions in the immunogenic compositions are used when chemically or heat treated mutated *Bordetella* strains are used in the vaccines or immunogenic compositions.

The immunogenic compositions of the invention further comprise at least one molecule having a prophylactic effect against a *Bordetella* infection or the detrimental effects of *Bordetella* infection, such as a nucleic acid, a protein, a polypeptide, a vector or a drug.

The immunogenic composition of the invention is used to elicit a T-cell immune response in a host in which the composition is administered. All immunogenic compositions described above can be injected in a host via different routes: subcutaneous (s.c.), intradermal (i.d.), intramuscular (i.m.) or intravenous (i.v.) injection, oral administration and intranasal administration or inhalation.

When formulated for subcutaneous injection, the immunogenic composition or vaccine of the invention preferably comprises between 10 and 100 µg of the *Bordetella* strain per injection dose, more preferably from 20 to 60 µg/dose, especially around 50 µg/dose, in a sole injection.

When formulated for intranasal administration, the *Bordetella* strain is administered at a dose of approximately  $1 \times 10^3$  to  $1 \times 10^6$  bacteria, depending on the weight and age of the mammal receiving it. In another aspect a dose of  $1 \times 10^4$  to  $5 \times 10^6$  can be used.

The mutated *Bordetella* strains of the present invention can be used as an attenuated vaccine to protect against future *Bordetella* infection. In this regard, an advantage of the present invention is that a single dose can be administered to mammals and the protection can last at least for a duration of longer than two months, particularly longer than six months. The vaccine of the present invention can be administered to newborns and protects against infection of whooping cough. This is especially crucial since the fatality rate from *Bordetella pertussis* infections is about 1.3% for infants younger than 1 month.

Moreover, the vaccines of the present invention can be used in adult mammals when there is an epidemic or in older adults over the age of 60, since their risk of complications maybe higher than that of older children or healthy adults.

The vaccines can be formulated with the physiological excipients set forth above in the same manner as in the immunogenic compositions. For instance, the pharmaceutically acceptable vehicles include, but are not limited to, phosphate buffered saline solutions, distilled water, emulsions such as an oil/water emulsions, various types of wetting agents sterile solutions and the like. Adjuvants such as muramyl peptides such as MDP, IL-12, aluminium phosphate, aluminium hydroxide, Alum and/or Montanide® can be used in the vaccines.

The vaccines of the present invention are able to induce high titers of serum IgG against FHA. The analysis of the antigen-specific cytokine patterns revealed that administration with the mutated attenuated *Bordetella* strains of the present invention favored a strong TH1 response.

The vaccines of the present invention provide high level of protection against a *Bordetella* infection i.e., a level of protection higher than 90%, particularly higher than 95%, more particularly higher than 99% (calculated 7 days after infection as detailed on example 9). The level of protection of the vaccine comprising the BPZE1 strain reaches more than 99.999% compared to non-vaccinated (naive) mice, at least two months after vaccination.

The vaccines can be administered subcutaneous (s.c.), intradermal (i.d.), intramuscular (i.m.) or intravenous (i.v.) injection, oral administration and intranasal administration or inhalation. The administration of the vaccine is usually in a single dose. Alternatively, the administration of the vaccine of the invention is made a first time (initial vaccination), followed by at least one recall (subsequent administration), with the same strain, composition or vaccine, or with acellular vaccines, or a combination of both.

In one aspect, intranasal administration or inhalation of the vaccines is accomplished, which type of administration is low in costs and enables the colonization by the attenuated strains of the invention of the respiratory tract: the upper respiratory tract nose and nasal passages, paranasal sinuses, and throat or pharynx) and/or the respiratory airways (voice box or larynx, trachea, bronchi, and bronchioles) and/or the lungs (respiratory bronchioles, alveolar ducts, alveolar sacs, and alveoli)

Intranasal administration is accomplished with an immunogenic composition or a vaccine under the form of liquid solution, suspension, emulsion, liposome, a cream, a gel or similar such multiphasic composition. Solutions and suspensions are administered as drops. Solutions can also be administered as a fine mist from a nasal spray bottle or from a nasal inhaler. Gels are dispensed in small syringes containing the required dosage for one application.

Inhalation is accomplished with an immunogenic composition or a vaccine under the form of solutions, suspensions, and powders; these formulations are administered via an aerosol or a dry powder inhaler. Compounded powders are administered with insufflators or puffers.

Use of the mutated *Bordetella* strains comprising at least a mutated ptx gene, a deleted or mutated dnt gene and a heterologous ampG gene for the preparation of a multivalent vaccine to treat respiratory diseases is yet another aspect of the present invention. In this regard, the attenuated mutated *Bordetella* strain described above, can be used as a heterologous expression platform to carry heterologous antigens to the respiratory mucosa. Thus, such respiratory pathogens such as *Neisseria*, *Pneumophila*, *yersinia*, *pseudomonas*, *mycobacteria*, influenza and the like can prevent infection using the BPZE1 as a carrier.

Use of the live attenuated mutated *Bordetella* strains described herein for the manufacture of a vaccine for the treatment or prevention of *Bordetella* infection is also encom-

passed by the present invention. In this regard, the vaccine can be used for the simultaneous treatment or prevention of an infection by *B. pertussis* and *B. parapertussis*.

Use of the vaccine to provide rapid protective immunity in case of a *pertussis* epidemic is also encompassed by the present invention.

Use of the vaccine to provide a rapid protective immunity, increasing over the at least next two months following vaccination is also encompassed by the present invention.

The vaccine or immunogenic composition is also provided in a kit. The kit comprises the vaccine or immunogenic composition and an information leaflet providing instructions for immunization.

The present invention also relates to a method for inducing T-cell mediated immune response and particularly a CD4<sup>+</sup>-mediated immune response or a CD8<sup>+</sup>-mediated immune response, comprising administering the live attenuated *Bordetella* strains of the invention in a non-human mammal or a human mammal.

A method of protecting a mammal against disease caused by infection by *Bordetella* comprising administering to said mammal in need of such treatment a mutated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene is another embodiment of the present invention. This method encompasses treating or preventing infections against *Bordetella pertussis* and/or *Bordetella parapertussis*. In one aspect the BPZE1 strain is used in this method.

Also a method of providing a rapid protective immunity against a *Bordetella* infection comprising administering to said mammal in need of such treatment a mutated *Bordetella* strain comprising at least a mutated ptx gene, a deleted or mutated dnt gene, and a heterologous ampG gene is encompassed by the present invention. In one aspect the BPZE1 strain is used in this method.

Moreover, the mutated live attenuated *Bordetella* strains of the present invention induce mucosal immunity, as well as systemic immunity. Thus, in another aspect the invention also relates to a method of inducing mucosal and systemic immunity by administering to a mammal in need of such treatment the mutated live attenuated *Bordetella* strains of the present invention. In one aspect the BPZE1 strain is used in this method.

Besides its role in the prevention and/or treatment of *Bordetella* infection, the mutated strain of the invention may be used as vector, to bear at least one further heterologous nucleic acid sequence encoding a RNA (such as antisense RNA) or a protein of interest. This means that the mutated strain bears at least one further heterologous nucleic acid sequence in addition to the heterologous ampG gene. In one aspect, the protein encoded by this at least one further heterologous nucleic acid sequence is a protein for which the expression is desired in the respiratory tract. In another aspect, the protein of interest is an antigen, such as a viral, a bacterial or a tumoral antigen, against which an immune response is desired. Therefore, the mutated *Bordetella* strain bearing at least one further heterologous nucleic acid sequence may also be used as a vaccine. The definitions given above for administration of the vaccine or immunogenic composition also apply to a vaccine comprising mutated *Bordetella* strain bearing at least one further heterologous nucleic acid sequence. Examples of heterologous proteins are antigens of pathogens causing infections of or diseases associated with the respiratory track: poliomyelitis, influenza (influenzavirus from Orthomyxoviridae family) or antigens from pneumococcus (such as *Streptococcus pneumoniae*).

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A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications may be made without departing from the spirit and scope of the invention.

## EXAMPLES

## Materials and Methods

## Example 1

*Bordetella* Strains and Growth Conditions

The *B. pertussis* strains used in this study were all derived from *B. pertussis* BPSM [13], and *B. parapertussis* is a streptomycin-resistant derivative of strain 12822 (kindly provided by Dr. N. Guiso, Institut Pasteur Paris, France). All *Bordetella* strains were grown on Bordet-Gengou (BG) agar (Difco, Detroit, Mich.) supplemented with 1% glycerol, 20% defibrinated sheep blood, and 100 µg/ml streptomycin. For cell adherence assays, exponentially growing *B. pertussis* was inoculated at an optical density of 0.15 at 600 nm in 2.5 ml modified Stainer-Scholte medium [14] containing 1 g/l heptakis(2,6-di-o-methyl) β-cyclodextrin (Sigma) and supplemented with 65 µCi/mi L-[<sup>35</sup>S]methionine plus L-[<sup>35</sup>S]cysteine (NEN, Boston, Mass.) and grown for 24 h at 37° C. The bacteria were then harvested by centrifugation, washed three times in phosphate-buffered saline (PBS) and resuspended in RPMI 1640 (Gibco. (Grand Island, N.Y.) at the desired density.

## Example 2

Construction of *B. pertussis* BPZE1

To construct *B. pertussis* BPZE1, the *B. pertussis* ampG gene was replaced by *Escherichia coli* ampG using allelic exchange. A PCR fragment named met and located at position 49,149 to 49,990 of the *B. pertussis* genome ([http://www.sanger.ac.uk/Projects/B\\_pertussis/](http://www.sanger.ac.uk/Projects/B_pertussis/)), upstream of the *B. pertussis* ampG gene, was amplified using oligonucleotides A: 5'-TATAAATCGATATTCCTGCTGGTTTCGTTCTC-3' (SEQ ID No:5) and B: 5'-TATAGCTAGCAAGTTGGGAAACGACACCAC-3' (SEQ ID No:6), and *B. pertussis* BPSM [13] genomic DNA as a template. This 634 bp fragment was inserted into Topo PCRII (InVitrogen Life Technology, Groningen, The Netherlands) and then excised as a ClaI-NheI fragment and inserted into ClaI- and NheI-digested pBP23 [50], a suicide vector containing the *E. coli* ampG gene with flanking *B. pertussis* DNA of 618 bp (from position 50,474 to 51,092 of the *B. pertussis* genome) and 379 bp (from position 52,581 to 52,960 of the *B. pertussis* genome) at the 5' and 3' end of *E. coli* ampG, respectively. The resulting plasmid was transferred into *E. coli* SM10 [51], which was then conjugated with BPSM, and two successive homologous recombination events were selected as described [52]. Ten individual colonies were screened by PCR as follows. The colonies were suspended in 100 µl H<sub>2</sub>O, heated for 20 min. at 95° C., and centrifuged for 5 min at 15,000×g. One µl of supernatants was then used as template for PCR using oligonucleotides A and C: 5'-TAAGAAGCAAAATAAGC-CAGGCATT-3' (SEQ ID No:7) to verify the presence of *E. coli* ampG and using oligonucleotides D: 5'-TATACCATG-GCGCCGCTGCTGGTGCTGGGC-3' (SEQ ID No:8) and E: 5'-TATATCTAGACGCTGGCCGTAACCTTAGCA-3' (SEQ ID No:9) to verify the absence of *B. pertussis* ampG. One of the strains containing *E. coli* ampG and lacking *B.*

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*pertussis* ampG was then selected, and the entire ampG locus was sequenced. This strain was then used for further engineering.

The ptx genes were deleted from the chromosome of this strain as described [21] and then replaced by mutated ptx coding inactive PTX. The EcoRI fragment containing the mutated ptx locus from pPT-RE [16] was inserted into the EcoRI site of pJQ200mp18rpsI [53]. The resulting plasmid was integrated into the *B. pertussis* chromosome at the ptx locus by homologous recombination after conjugation via *E. coli* SM10. The ptx locus in the chromosome of the resulting *B. pertussis* strain was sequenced to confirm the presence of the desired mutations. Toxin production was analyzed by immunoblotting using a mix of monoclonal antibodies IB7 [54] specific for subunit S1 and 11E6 [55] specific for subunits S2 and S3 of PTX.

Finally, the dnt gene was deleted from the resulting *B. pertussis* strain as the dnt flanking regions were amplified by PCR using BPSM genomic DNA as a template and oligonucleotides F: 5'-TATAGAATTCGCTCGGTTTCGCTGGT-CAAG G-3' (SEQ ID No: 10) and G: 5'-TATATCTAGAG-CAATGCCGATTCATCTTTA-3' (SEQ ID No: 11) for the dnt upstream region, and H: 5'-TATATCTAGAGCGCCTT TATTGCTTTTCC-3' (SEQ ID No: 12) and I: 5'-TATAAAGCTTCTCATGCACGCCG GCTTCTC-3' (SEQ ID No:13) for the dnt downstream region, as primers. The resulting 799-bp and 712-bp DNA fragments were digested with EcoRI/XbaI and XbaI/HindIII, respectively, and linked together using the Fast Link kit (Epicentre Biotechnologies, Madison, Wis.). The ligated fragment was amplified by PCR using oligonucleotides F and I, and the 1505-bp PCR fragment was then inserted into pCR2.1-Topo (Invitrogen), re-isolated from the resulting plasmid as an EcoRI fragment and inserted into the unique EcoRI site of pJQmp200rpsI.18. The resulting plasmid was introduced into *B. pertussis* by conjugation via *E. coli* SM10. Successful deletion of the dnt gene by allelic exchange was verified by Southern blot analysis on PvuII-digested *B. pertussis* genomic DNA using the PCR fragment corresponding to the dnt upstream region as a probe. The probe was labeled with digoxigenin (DIG) using the DIG Easy Hyb labeling kit. (Roche, Meylan, France). The sizes of the hybridizing bands were determined from the migration distance of the Dig-labeled DNA molecular marker III (Roche). The dnt locus of this final strain, named BPZE1 was sequenced.

## Example 3

## Analysis of TCT Production

For sensitive quantitation of TCT production, culture supernatants of *B. pertussis* grown to logarithmic phase were collected, subjected to solid phase extraction [15] and derivatized with phenylisothiocyanate (PITC, Pierce). The resulting phenylthiocarbonyl (PTC) derivatives were separated by reversed-phase HPLC using a C8 column (Perkin Elmer) and detected at 254 nm. The amount of *B. pertussis* PTC-TCT in each sample was determined by comparing the peak area and elution time with an identically processed TCT standard.

## Example 4

## Cell-Adherence Assay

To analyze adherence properties of the *B. pertussis* strains, their attachment rates to the human pulmonary epithelial cell

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line A549 (ATCC n° CCL-185) and the murine macrophage cell line J774 (ATCC n° TIB-67) were measured as previously described [16].

## Example 5

## Transmission Electron Microscopy

The single droplet-negative staining procedure was used as described previously [17] with the following modifications. 20 µl of a suspension at approximately 10<sup>9</sup> bacteria/ml were absorbed for 2 min. onto form forward carbon-coated nickel grids (400 mesh; Electron Microscopy Sciences EMS, Washington, Pa.). After 30 seconds air-drying the grids were stained for 2 minutes with 20 µl of 2% phosphotungstic acid (pH17; EMS) and examined after air-drying under a transmission electron microscope (Hitachi 7500, Japan) at 60 kvolts and high resolution.

## Example 6

## Intranasal Infection and Vaccination

3-week and 8-week old female Balb/C were kept under specific pathogen-free conditions, and all experiments were carried out under the guidelines of the Institut Pasteur de Lille animal study board. Mice were intranasally infected with approximately 4×10<sup>6</sup> bacteria in 20 µl PBS, and kinetics of CFU in the lungs were measured as previously described [18]. For vaccination with aPV (Tetravac; Aventis-Pasteur, France), mice were immunized intraperitoneally (i.p.) with 20% of the human dose and boosted one month later using the same dose.

## Example 7

## Antibody Determination

Sera were collected, and antibody titers were estimated by enzyme-linked immunosorbent assays (ELISA) as previously described [18].

## Example 8

## Cytokine Assays

Spleen cells from individual mice were tested at different time points after immunization for in vitro cytokine production in response to heat-killed *B. pertussis* BPSM (10<sup>6</sup> cells/ml), 5.0 µg/ml PTX (purified from *B. pertussis* BPGR4 [19] as previously described [20] and heat-inactivated at 80° C. for 20 min). 5.0 µg filamentous hemagglutinin (FHA, purified from *B. pertussis* BPRA [21] as previously described [22]), 5 µg/ml concanavalin A (Sigma Chemical Co., St. Louis, Mo.) or medium alone as control. Supernatants were removed from triplicate cultures after 72 h incubation at 37° C. and 5% (CO<sub>2</sub>, and IFN-γ and IL-5 concentrations were determined by immunoassays (BD OptEIA set, Pharmingen).

## Example 9

## Intranasal Infection and Vaccination: Challenge at 1, 2, 3 and 4 Weeks

An infant (3 weeks-old) mouse model [29] was used to compare the efficiency of vaccination with BPZE1 with the one of vaccination with acellular *pertussis* vaccine (aPv).

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Female Balb/C mice were intranasally infected with approximately 1×10<sup>6</sup> BPZE1 strain in 20 µl PBS. For vaccination with aPv (Tetravac; Aventis-Pasteur, France), mice were immunized intraperitoneally with 20% of the human dose. One, two, three or four weeks after vaccination with BPZE1 or aPv, mice were intranasally challenged with virulent *B. pertussis* BPSM/betA-lacZ strain [53]. This strain is a BPSM-derivative gentamycin-resistant which allows the discrimination with BPZE1 (gentamycin-sensitive) on Bordet-Gengou agar plates containing 10 µg/ml of gentamycin and 100 µg/ml of streptomycin (BGgs). Control group corresponds to naive mice challenged with BPSM/betA-lacZ. One week after challenge infection, lungs were aseptically removed, homogenized and plates on BGgs for CFU determination as previously described [18].

Mice were vaccinated with BPZE1 or aPv and challenged with virulent *B. pertussis* one, two, three or four weeks after vaccination. Lung CFUs counts were determined 3 hours or 7 days later. Results are expressed as mean (±standard error) CFUs from three to five mice per group. Levels of protection are calculated for each challenge infection as mean percentages of CFUs of each group relative of the average of CFUs in non-immunized group, 7 days after challenge infection (Tables 2 to 5).

## Example 10

## Statistical Analysis

The results were analyzed using the unpaired Student's t test and the Kruskal-Wallis test followed by the Dunn's post-test (GraphPad Prism program) when appropriate. Differences were considered significant at P≤0.05.

## Results

35 Construction of *B. pertussis* BPZE1

Three virulence factors were genetically targeted: tracheal cytotoxin (TCT), *pertussis* toxin (PTX) and dermonecrotic toxin (DNT).

TCT is responsible for the destruction of ciliated cells in the trachea of infected hosts [24, 25] and may thus be involved in the cough syndrome. TCT is a breakdown product of peptidoglycan in the cell wall of Gram-negative bacteria, which generally internalize it into the cytosol by the AmpG transporter protein to be re-utilized during cell wall biosynthesis. *B. pertussis* AmpG is inefficient in the internalization of peptidoglycan breakdown products. We therefore replaced the *B. pertussis* ampG gene by *E. coli* ampG. The resulting strain expressed less than 1% residual TCT activity (FIG. 1).

PTX is a major virulence factor responsible for the systemic effects of *B. pertussis* infections and is composed of an enzymatically active moiety, called S1, and a moiety responsible for binding to target cell receptors (for review, see 26). However, it is also one of the major protective antigens, which has prompted us to replace the natural ptx genes by a mutated version coding for an enzymatically inactive toxin. This was achieved by replacing Lys-9 by Arg and Glu-129 by Gly in S1, two key residues involved in substrate binding and catalysis, respectively. Allelic exchange was used to first delete the ptx operon, and then to insert the mutated version. The presence of the relevant toxin analogues in the *B. pertussis* culture supernatants was evaluated by immunoblot analysis (FIG. 2).

Finally, allelic exchange was used to remove the dnt gene (FIG. 3). Although the role of DNT in the virulence of *B. pertussis* is not certain, it has been identified as an important toxin in the closely related species *Bordetella bronchiseptica* and displays lethal activity upon injection of minute quantities (for review, see 26).

#### In Vitro Characterization of *B. pertussis* BPZE1

Since some of the genetic alterations in BPZE1 may potentially affect the bacterial cell wall synthesis, the size and shape, as well as the in vitro growth rate of BPZE1 was compared with those of the parental strain BPSM. The growth rate of BPZE1 did not differ from that of BPSM (FIG. 4), and no difference in bacterial shape or size was detected between BPZE1 and BPSM, as evidenced by electron microscopy analysis (FIG. 5). However, the cell wall of BPZE1 appeared to be consistently somewhat thinner than that of BPSM.

To determine whether the absence or alterations of any of the targeted toxins in BPZE1 affects adherence properties of *B. pertussis*, the attachment rates of BPZE1 was compared with those of BPSM, using the human pulmonary epithelial cell line A549 and the murine macrophage cell line J774, as two cellular models often used to study the adherence of *B. pertussis*. No significant difference in the adherence capacities to either cell line was observed between the two strains (FIG. 6).

#### Attenuation of *B. pertussis* BPZE1

To determine whether the mutations introduced into *B. pertussis* BPZE1 have resulted in attenuation, yet allow the organism to colonize the respiratory tract, Balb/C mice were intranasally infected with BPZE1 or BPSM, and colonization was followed over time. BPZE1 was able to colonize and persist in the lungs of mice as long as BPSM (FIG. 7). However, the peak of multiplication seen 7 days after infection with BPSM was consistently lacking in mice infected with BPZE1. Studies done with strains mutated in individual toxin genes indicated that this is due to the mutations in the *ptx* locus (data not shown). When the lungs were examined for histopathological changes and inflammatory infiltration, infection with BPSM was found to induce strong peri-bronchiovascular infiltrates and inflammatory cell recruitment 7 days after infection, associated with a strong hypertrophy of the bronchiolar epithelial cells (FIG. 8). In contrast, no such changes were seen in BPZE1-infected animals, and the histology of the BPZE1-infected mice was similar to that of the control mice that had received PBS instead of the bacteria. The BPSM-infection induced inflammation lasted for at least two months (data not shown). These results indicate that the mutations introduced into BPZE1 have resulted in drastic attenuation, but allow the bacteria to colonize and persist in the lungs.

#### Protection Against *B. pertussis* Challenge after Intranasal Vaccination of Adult Mice with BPZE1

To evaluate the protection offered by BPZE1, the effect of a single intranasal administration of this strain to 8-weeks old Balb/C mice on the subsequent colonization by the wild type challenge strain BPSM was compared with that of two i. p. immunizations with 1/5 of a human dose of aPV. This aPV immunization protocol has been described as the best to correlate with *pertussis* vaccine efficacy in human clinical trials [27, 28]. As shown by the total clearance of bacterial colony counts in the lungs seven days after challenge infection, a single intranasal administration of BPZE1 and two i.p. immunizations with aPV provided similar levels of protection (FIG. 9a). High bacterial loads were found in the control mice that had received two injections of PBS instead of the vaccine.

#### Protection Against *B. pertussis* Challenge after Intranasal Vaccination of Infant Mice with BPZE1

Since the principal targets of novel *pertussis* vaccines are young infants, that are not protected with the currently available vaccines, an infant (3 weeks-old) mouse model [29] was developed and used to compare the efficiency of vaccination with BPZE1 with that of vaccination with aPV. A single nasal administration of BPZE1 fully protected infant mice against

challenge infection (FIG. 9b), as complete bacterial clearance was observed in the lungs one week after challenge. In contrast, substantial numbers of bacteria remained in the aPV-vaccinated animals one week after challenge infection. The difference in bacterial load between the BPZE1-vaccinated and the aPV-vaccinated mice was statistically significant, indicating that in the infant mouse model a single intranasal administration with BPZE1 provides better protection than two systemic administrations of aPV.

In addition, a strong reduction in the bacterial load of the challenge strain 3 hours after administration when the mice had been immunized with BPZE1 was consistently observed compared to the aPV-immunized animals (FIG. 9c), indicating that vaccination with BPZE1 reduces the susceptibility to infection by the challenge strain. This effect was seen in both 8-weeks old and in infant mice. In contrast, aPV had no effect on the bacterial counts 3 hours after infection, when compared to the control mice.

#### Protection Against *B. parapertussis* Challenge after Intranasal Vaccination with BPZE1

There is increasing concern about *B. parapertussis* infection in children, especially in immunized populations [30, 31]. *B. parapertussis* causes a milder *pertussis*-like syndrome, the frequency of which is probably largely underestimated. Furthermore, the incidence of *B. parapertussis* infections has been increasing over the last decades, possibly due to the fact that *pertussis* vaccines are known to have very low or no protective efficacy against *B. parapertussis* [32, 33]. In contrast, infection by *B. pertussis* has recently been reported to protect against *B. parapertussis* infection [34]. BPZE1 was also assessed for protection against *B. parapertussis* using the infant mouse model. Whereas two administrations of aPV did not provide any protection against *B. parapertussis*, as previously reported, a single intranasal administration of BPZE1 provided strong protection, as measured by the low numbers of *B. parapertussis* counts in the lungs of the vaccinated mice 1 week after challenge (FIG. 9d).

#### Immune Responses Induced by BPZE1 Vaccination

Although the mechanisms of protective immunity against *B. pertussis* infection are not yet completely understood, clear evidence of a role for both B cells and IFN- $\gamma$  has been demonstrated in mice [28]. Vaccination with either one nasal dose of BPZE1 or two i. p. administrations of aPV induced high titers of serum IgG against FHA, a major surface antigen of *B. pertussis* [35], also present in aPV (FIG. 10a). Following *B. pertussis* challenge, positive anamnestic responses were measured in BPZE1- and in aPV-vaccinated animals, as indicated by an increase in anti-FHA IgG titers, compared to primary responses before *B. pertussis* infection. Examination of the anti-FHA IgG1/IgG2a ratios showed that these ratios were higher after aPV administration, characteristic of a Th2 type response, than after BPZE1 vaccination (FIG. 10b). Although the anti-FHA-IgG1/IgG2a decreased after challenge in the aPV vaccinated mice, it remained still substantially higher than in the BPZE1-vaccinated animals after *B. pertussis* challenge.

Analysis of *B. pertussis* antigen-specific cytokine patterns induced by BPZE1 or aPV vaccination confirmed that BPZE1 administration favors a stronger Th1 type response than aPV vaccination. This was revealed by the fact that the ratios of IFN- $\gamma$  over IL-5 produced by splenocytes stimulated with FHA or PT, or with the polyclonal activator ConA were significantly higher in BPZE1 vaccinated mice than in aPV vaccinated mice (FIG. 10c).

Protective Immunity of BPZE1 Over Time (from 1 Week to 4 Weeks)

As shown in Tables 1 to 5 below, whereas administration of aPv provided limited protection (reduction of 75% of bacterial load compared to non-vaccinated mice at 1 week) against *B. pertussis*, a single intranasal administration of BPZE1 already provided high level of protection (reduction of 97.64% of bacterial load) against a *B. pertussis* challenge infection performed one week after vaccination. If challenge

infection occurred two weeks after vaccination, the level of protection induced by BPZE1 reached more than 99.999% compared to non-vaccinated mice and is significantly superior to the protection induced by aPv vaccine (approximately 92% compared to non-vaccinated mice). Therefore, vaccine efficacy of BPZE1 against *B. pertussis* challenge is already significant one week after vaccination and is increasing over the at least next two months.

TABLE 1

Kinetics of vaccines efficacy against <i>B. pertussis</i> challenge in infant mice.				
Time between vaccination and challenge	Time between lungs recovery and challenge	Log10 cfu/lungs of mice		
		Naive	aPv-vaccinated	BPZE1-vaccinated
1 week	3 hours	5.71 ± 0.03	5.8 ± 0.07	5.74 ± 0.01
	7 days	6.71 ± 0.06	5.97 ± 0.20	4.86 ± 0.35
2 weeks	3 hours	5.77 ± 0.10	5.60 ± 0.02	5.49 ± 0.05
	7 days	6.49 ± 0.08	5.31 ± 0.16	3.22 ± 0.33
3 weeks	3 hours	6.03 ± 0.11	5.88 ± 0.04	5.33 ± 0.08
	7 days	6.58 ± 0.09	5.62 ± 0.11	3.14 ± 0.38
4 weeks	3 hours	6.31 ± 0.01	6.15 ± 0.02	5.83 ± 0.05
	7 days	6.36 ± 0.04	5.21 ± 0.11	1.83 ± 0.46

TABLE 2

Level of protection of aPv-vaccinated and BPZE1-vaccinated mice as compared to non-vaccinated mice at week 1.				
Non vaccinated mice	Number of bacteria in lungs		Mean number of bacteria	
Non-vaccinated 1	4.7 × 10 <sup>6</sup>		5.36 · 10 <sup>6</sup>	
Non-vaccinated 2	3.8 × 10 <sup>6</sup>			
Non-vaccinated 3	8.2 × 10 <sup>6</sup>			
Non-vaccinated 4	4.1 × 10 <sup>6</sup>			
Non-vaccinated 5	6 × 10 <sup>6</sup>			
	Number of bacteria in lungs	Percentage of remaining bacteria <sup>(1)</sup>	Mean percentage of remaining bacteria	Level of protection
aPv-vaccinated mice				
aPv1	1.95 × 10 <sup>6</sup>	36.38	25%	75%
aPv2	2.9 × 10 <sup>6</sup>	54.1		
aPv3	2.9 × 10 <sup>5</sup>	5.41		
aPv4	3.6 × 10 <sup>5</sup>	6.72		
aPv5	1.2 × 10 <sup>6</sup>	22.39		
BPZE1-vaccinated mice				
BPZE1-1	3.2 × 10 <sup>5</sup>	5.97	2.36%	97.64%
BPZE1-2	2 × 10 <sup>4</sup>	0.004		
BPZE1-3	6 × 10 <sup>4</sup>	1.12		

<sup>(1)</sup> Percentage of remaining bacteria = number of bacteria for each particular mouse/mean number of bacteria of all non-vaccinated mice

TABLE 3

Level of protection of aPv-vaccinated and BPZE1-vaccinated mice as compared to non-vaccinated mice at week 2.		
Non vaccinated mice	Number of bacteria in lungs	Mean number of bacteria
Non-vaccinated 1	5 × 10 <sup>6</sup>	3.34 × 10 <sup>6</sup>
Non-vaccinated 2	3.6 × 10 <sup>6</sup>	
Non-vaccinated 3	1.7 × 10 <sup>6</sup>	
Non-vaccinated 4	2.4 × 10 <sup>6</sup>	
Non-vaccinated 5	4 × 10 <sup>6</sup>	



TABLE 3-continued

Level of protection of aPv-vaccinated and BPZE1-vaccinated mice as compared to non-vaccinated mice at week 2.				
	Number of bacteria in lungs	Percentage of remaining bacteria <sup>(1)</sup>	Mean percentage of remaining bacteria	Level of protection
aPv-vaccinated mice				
aPv1	$9.5 \times 10^4$	2.84	8.11%	91.89%
aPv2	$2.9 \times 10^5$	8.68		
aPv3	$1 \times 10^5$	2.99		
aPv4	$6.8 \times 10^5$	20.36		
aPv5	$1.9 \times 10^5$	5.69		
BPZE1-vaccinated mice				
BPZE1-1	$9.5 \times 10^3$	$2.8 \times 10^{-3}$	$1.03 \times 10^{-3}\%$	99.999%
BPZE1-2	450	$1.35 \times 10^{-4}$		
BPZE1-3	3500	$1.05 \times 10^{-3}$		
BPZE1-4	500	$1.5 \times 10^{-4}$		

<sup>(1)</sup> Percentage of remaining bacteria = number of bacteria for each particular mouse/mean number of bacteria of all non-vaccinated mice

TABLE 4

Level of protection of aPv-vaccinated and BPZE1-vaccinated mice as compared to non-vaccinated mice at week 3.		
Non vaccinated mice	Number of bacteria in lungs	Mean number of bacteria
Non-vaccinated 1	$1.8 \times 10^6$	$4.04 \times 10^6$
Non-vaccinated 2	$5.75 \times 10^6$	
Non-vaccinated 3	$4.7 \times 10^6$	
Non-vaccinated 4	$3.2 \times 10^6$	
Non-vaccinated 5	$4.75 \times 10^6$	

	Number of bacteria in lungs	Percentage of remaining bacteria <sup>(1)</sup>	Mean percentage of remaining bacteria	Level of protection
aPv-vaccinated mice				
aPv1	$1.99 \times 10^5$	4.94	11.26%	88.74%
aPv2	$6 \times 10^5$	14.85		
aPv3	$6 \times 10^5$	14.85		
aPv4	$4.2 \times 10^5$	10.40		
BPZE1-vaccinated mice				
BPZE1-1	3640	$9.01 \times 10^{-4}$	$8.65 \times 10^{-4}\%$	99.999%
BPZE1-2	9720	$2.4 \times 10^{-3}$		
BPZE1-3	300	$7.43 \times 10^{-5}$		
BPZE1-4	340	$8.42 \times 10^{-5}$		

<sup>(1)</sup> Percentage of remaining bacteria = number of bacteria for each particular mouse/mean number of bacteria of all non-vaccinated mice

TABLE 5

Level of protection of aPv-vaccinated and BPZE1-vaccinated mice as compared to non-vaccinated mice at week 4.		
Non vaccinated mice	Number of bacteria in lungs	Mean number of bacteria
Non-vaccinated 1	$2.1 \times 10^6$	$2.36 \times 10^6$
Non-vaccinated 2	$2.2 \times 10^6$	
Non-vaccinated 3	$3.1 \times 10^6$	
Non-vaccinated 4	$2.6 \times 10^6$	
Non-vaccinated 5	$1.8 \times 10^6$	

	Number of bacteria in lungs	Percentage of remaining bacteria <sup>(1)</sup>	Mean percentage of remaining bacteria	Level of protection
aPv-vaccinated mice				
aPv1	$2.52 \times 10^5$	10.68	7.76%	92.24%
aPv2	$3.28 \times 10^5$	13.90		
aPv3	$1.04 \times 10^5$	4.41		
aPv4	$8.4 \times 10^5$	3.56		
aPv5	$1.48 \times 10^5$	6.27		

TABLE 5-continued

Level of protection of aPV-vaccinated and BPZE1-vaccinated mice as compared to non-vaccinated mice at week 4.				
BPZE1-vaccinated mice				
BPZE1-1	190	$8.05 \times 10^{-5}$	$7.13 \times 10^{-5}\%$	99.999%
BPZE1-2	0	0		
BPZE1-3	110	$4.66 \times 10^{-5}$		
BPZE1-4	320	$1.36 \times 10^{-4}$		
BPZE1-5	220	$9.32 \times 10^{-5}$		

<sup>(1)</sup> Percentage of remaining bacteria = number of bacteria for each particular mouse/mean number of bacteria of all non-vaccinated mice

## DISCUSSION

*Pertussis* is the first infectious disease whose incidence is increasing in countries with high vaccine coverage. This paradoxical situation is most likely linked to the epidemiological changes observed since the massive introduction of highly efficacious vaccines. In contrast to the pre-vaccination era, cases of adolescent and adult *pertussis* are now increasingly more frequent. Although generally not life-threatening in that age group, *B. pertussis*-infected adults are an important reservoir for infection of the very young children, too young to be protected by vaccination. Early vaccination, possibly at birth, would therefore be highly desirable, but is hampered by the immaturity of the immune system of neonates and infants. However, the fact that natural *B. pertussis* infection, even very early in life, is able to induce a strong Th1 response in infants [12] prompted us to develop a live attenuated *B. pertussis* vaccine strain to be given by the nasal route as an alternative over the currently available vaccines.

Based on experimental infections of primates, Huang et al. had already in 1962 come to the conclusion that ultimate protection against whooping cough probably best follows a live *B. pertussis* inoculation [36]. In veterinary medicine, attenuated *Bordetella* strains have been used to vaccinate against bordetellosis in dogs and piglets. A live attenuated *Bordetella bronchiseptica* strain has been shown to provide strong protection against kennel cough in dogs [37] after nasal administration. This protection was seen as early as 48 h after vaccination. Intranasal vaccination with live attenuated *B. bronchiseptica* has also been shown to protect against atrophic rhinitis in two-days old piglets [38], indicating that in a live attenuated form *Bordetella* vaccines can be highly active in new-born animals.

Previous attempts to genetically attenuate *B. pertussis* as a live vaccine candidate have met with rather limited success. Based on a strategy used for the successful attenuation of *Salmonella* vaccine strains [39], Roberts et al. have deleted the *aroA* gene of *B. pertussis* [40]. The *aroA* mutant was indeed highly attenuated, but had also lost its capacity to colonize the respiratory tract of the intranasally vaccinated animals and induced protective immunity only after repeated administrations of high doses. We took advantage of the knowledge on the molecular mechanisms of *B. pertussis* virulence and developed the highly attenuated strain BPZE1. This strain contains genetic alterations leading to the absence or inactivation of three major toxins, PTX, TCT and DNT. In contrast to the *aroA* mutant, this strain was able to colonize the mouse respiratory tract and to provide full protection after a single intranasal administration. The protection in adult mice was indistinguishable from that induced by two administrations of 1/5 of a human dose of aPV. An important difference, however, was seen in infant mice, where a single administration of BPZE1 fully protected, whereas aPV only offered

partial protection. In the context of the difficulties to induce protection in infants with the currently available vaccines early in life, these results provide hope for the development of novel vaccination strategies that may be used in the very young children, possibly at birth. In addition, BPZE1 protected against *B. parapertussis*, whereas aPV did not. Therefore the use of BPZE1 should also have an impact on the incidence of whooping cough caused by *B. parapertussis* in infants.

Although the recent replacement of first generation whole-cell vaccines by new aPV in many countries has significantly reduced the systemic adverse reactions observed with whole-cell vaccines, it has not abolished the need for repeated vaccination to achieve protection. This makes it unlikely to obtain protection in very young children (<6 months) that present the highest risk to develop severe disease. In addition, the wide-spread use of aPV has revealed new, unforeseen problems. Repeated administration of aPV may cause extensive swelling at the site of injection [41], which was not observed with whole-cell vaccines. In approximately 5% of the cases this swelling involves almost the entire limb and lasts for more than a week. Although the mechanism of this swelling has not been characterized yet, it has been proposed to be due to an Arthus hypersensitivity reaction caused by high antibody levels induced by the primary immunization [42]. However, it could also be related to the Th2 skewing of the immune response, as, compared to whole-cell vaccines, aPV administration induces more Th2-type cytokines in vaccinated children [10] and causes a delay in the Th1 development (Mascart et al., in preparation). Delayed maturation of Th1 function has been associated with a risk for atopy in genetically pre-disposed individuals [33]. The two mechanisms are not mutually exclusive. Compared to aPV, the immune response to BPZE1 administration is less biased towards the Th2 arm, and since BPZE1 is administered mucosally, no swelling reaction can occur.

The use of live attenuated bacteria as vaccines raises the issue of their biosafety. As such, they fall under the directives and guidelines for genetically modified organisms susceptible to be released into the environment. These guidelines and directives describe several objectives that have to be met, including hazard identification and environmental risk assessment [44]. Potential pathogenicity needs to be carefully considered, especially in immuno-compromised individuals, such as those infected with HIV. The natural biology of *B. pertussis* is particularly interesting in that regard. Although *pertussis* in HIV-infected subjects has been described occasionally, it is rather rare in AIDS patients [45]. In its genetically attenuated form, *B. pertussis* would therefore not be expected to cause major problems in HIV-infected children, especially if severe AIDS is an exclusion criterion, as it is for many vaccines. *B. pertussis* colonization is strictly limited to the respiratory epithelium, without extrapulmonary dissemi-

nation of the bacteria, which naturally excludes systemic bacteremia of the BPZE1 vaccine strain. If nevertheless unforeseeable safety problems occurred, the vaccine strain can easily be eliminated by the use of macrolide antibiotics, such as erythromycin, to which essentially all *B. pertussis* isolates are highly sensitive.

A further concern, like for any live vaccine, is the potential release of the vaccine strain in the environment and the consequences of such a release. *B. pertussis* is a strictly human pathogen, and there is no animal vector or reservoir. Moreover, unlike *B. bronchiseptica*, survival of wild-type *B. pertussis* in the environment is extremely limited [46]. *Pertussis* is only spread by coughing individuals, and there appears to be no asymptomatic carriage [47]. Coughing cannot be assessed in the mouse models used in this study. However, due to the nature of the genetic alterations in BPZE1, in particular the strong reduction of TCT and the genetic inactivation of PTX, this strain would not be expected to induce coughing. Active PTX has been shown to be required for cough induction in a coughing rat model, although the mechanism is not known [48]. If the vaccine strain were nevertheless to be transmitted to non-vaccinated individuals, this would at worst result in increased vaccine coverage. The consequences of each of these potential hazards can thus be graded as negligible and can easily and rapidly be controlled by antibiotic treatment if necessary.

Advantages of the use of BPZE1 include the relatively low production costs, making it especially attractive for developing countries, its needle-free easy and safe mode of administration and its potential to induce mucosal immunity in addition to systemic immunity. Although the role of mucosal immunity against *pertussis* has surprisingly not been much addressed, the fact that *B. pertussis* is a strictly mucosal pathogen, makes it likely that mucosal immune responses may contribute significantly to protection. None of the currently available vaccines induces any significant mucosal response.

Other advantages of the use of BPZE1 in vaccination are: the rapid protective immune response obtained after a single intranasal dose of BPZE1, since induction of the immunity can be detected 1 week after vaccination.

an increase of the protective immunity over the at least next two months after vaccination, and

the complete protective immunity, since a level of protection of more than 99.999% is obtained 2 weeks after vaccination.

The use of live attenuated *B. pertussis* for mucosal vaccination offers yet another advantage. *B. pertussis* can be used for the presentation of heterologous antigens to the respiratory mucosa (for review see 49). The use of BPZE1 as a heterologous expression platform may thus be helpful for the generation of multivalent vaccines against a variety of respiratory pathogens. However, since intranasal delivery of BPZE1 also induces strong systemic immune responses, as shown here by both the high levels of anti-FHA antibodies and of antigen-specific IFN- $\gamma$  production, it may also be used for the production of antigens to which systemic immune responses are desired.

While the invention has been described in terms of various preferred embodiments, the skilled artisan will appreciate that various modifications, substitutions, omissions and changes may be made without departing from the scope thereof. Accordingly, it is intended that the scope of the present invention be limited by the scope of the following claims, including equivalents thereof.

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## SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 13

<210> SEQ ID NO 1

<211> LENGTH: 269

<212> TYPE: PRT

<213> ORGANISM: *Bordetella pertussis*

<400> SEQUENCE: 1

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Met Arg Cys Thr Arg Ala Ile Arg Gln Thr Ala Arg Thr Gly Trp Leu
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Thr Trp Leu Ala Ile Leu Ala Val Thr Ala Pro Val Thr Ser Pro Ala
 20           25           30

Trp Ala Asp Asp Pro Pro Ala Thr Val Tyr Arg Tyr Asp Ser Arg Pro
 35           40           45

Pro Glu Asp Val Phe Gln Asn Gly Phe Thr Ala Trp Gly Asn Asn Asp
 50           55           60

Asn Val Leu Asp His Leu Thr Gly Arg Ser Cys Gln Val Gly Ser Ser
 65           70           75           80

Asn Ser Ala Phe Val Ser Thr Ser Ser Ser Arg Arg Tyr Thr Glu Val
 85           90           95

Tyr Leu Glu His Arg Met Gln Glu Ala Val Glu Ala Glu Arg Ala Gly
100           105           110

Arg Gly Thr Gly His Phe Ile Gly Tyr Ile Tyr Glu Val Arg Ala Asp
115           120           125

Asn Asn Phe Tyr Gly Ala Ala Ser Ser Tyr Phe Glu Tyr Val Asp Thr
130           135           140

Tyr Gly Asp Asn Ala Gly Arg Ile Leu Ala Gly Ala Leu Ala Thr Tyr
145           150           155           160

Gln Ser Glu Tyr Leu Ala His Arg Arg Ile Pro Pro Glu Asn Ile Arg
165           170           175

Arg Val Thr Arg Val Tyr His Asn Gly Ile Thr Gly Glu Thr Thr Thr
180           185           190

Thr Glu Tyr Ser Asn Ala Arg Tyr Val Ser Gln Gln Thr Arg Ala Asn
195           200           205

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Pro Asn Pro Tyr Thr Ser Arg Arg Ser Val Ala Ser Ile Val Gly Thr
 210                215                220

Leu Val Arg Met Ala Pro Val Ile Gly Ala Cys Met Ala Arg Gln Ala
225                230                235                240

Glu Ser Ser Glu Ala Met Ala Ala Trp Ser Glu Arg Ala Gly Glu Ala
                245                250                255

Met Val Leu Val Tyr Tyr Glu Ser Ile Ala Tyr Ser Phe
                260                265

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<210> SEQ ID NO 2
<211> LENGTH: 1464
<212> TYPE: PRT
<213> ORGANISM: Bordetella pertussis

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<400> SEQUENCE: 2

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Met Asp Lys Asp Glu Ser Ala Leu Arg Gln Leu Val Asp Met Ala Leu
 1          5          10          15

Val Gly Tyr Asp Gly Val Val Glu Glu Leu Leu Ala Leu Pro Ser Glu
 20          25          30

Glu Ser Gly Asp Leu Ala Gly Gly Arg Ala Lys Arg Glu Lys Ala Glu
 35          40          45

Phe Ala Leu Phe Ser Glu Ala Pro Asn Gly Asp Glu Pro Ile Gly Gln
 50          55          60

Asp Ala Arg Thr Trp Phe Tyr Phe Pro Lys Tyr Arg Pro Val Ala Val
 65          70          75          80

Ser Asn Leu Lys Lys Met Gln Val Ala Ile Arg Ala Arg Leu Glu Pro
 85          90          95

Glu Ser Leu Ile Leu Gln Trp Leu Ile Ala Leu Asp Val Tyr Leu Gly
100          105          110

Val Leu Ile Ala Ala Leu Ser Arg Thr Val Ile Ser Asp Leu Val Phe
115          120          125

Glu Tyr Val Lys Ala Arg Tyr Glu Ile Tyr Tyr Leu Leu Asn Arg Val
130          135          140

Pro His Pro Leu Ala Thr Ala Tyr Leu Lys Arg Arg Gln Arg Pro
145          150          155          160

Val Asp Arg Ser Gly Arg Leu Gly Ser Val Phe Glu His Pro Leu Trp
165          170          175

Phe Ala Tyr Asp Glu Leu Ala Gly Thr Val Asp Leu Asp Ala Asp Ile
180          185          190

Tyr Glu Gln Ala Leu Ala Glu Ser Ile Glu Arg Arg Met Asp Gly Glu
195          200          205

Pro Asp Asp Gly Ser Leu Asp Thr Ala Glu His Asp Val Trp Arg Leu
210          215          220

Cys Arg Asp Gly Ile Asn Arg Gly Glu Gln Ala Ile Phe Gln Ala Ser
225          230          235          240

Gly Pro Tyr Gly Val Val Ala Asp Ala Gly Tyr Met Arg Thr Val Ala
245          250          255

Asp Leu Ala Tyr Ala Asp Ala Leu Ala Asp Cys Leu His Ala Gln Leu
260          265          270

Arg Ile Arg Ala Gln Gly Ser Val Asp Ser Pro Gly Asp Glu Met Pro
275          280          285

Arg Lys Leu Asp Ala Trp Glu Ile Ala Lys Phe His Leu Ala Ala Thr
290          295          300

Gln Gln Ala Arg Val Asp Leu Leu Glu Ala Ala Phe Ala Leu Asp Tyr
305          310          315          320

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Ala Ala Leu Arg Asp Val Arg Val Tyr Gly Asp Tyr Arg Asn Ala Leu  
325 330 335

Ala Leu Arg Phe Ile Lys Arg Glu Ala Leu Arg Leu Leu Gly Ala Arg  
340 345 350

Arg Gly Asn Ala Ser Thr Met Pro Ala Val Ala Ala Gly Glu Tyr Asp  
355 360 365

Glu Ile Val Ala Ser Gly Ala Ala Asn Asp Ala Ala Tyr Val Ser Met  
370 375 380

Ala Ala Ala Leu Ile Ala Gly Val Leu Cys Asp Leu Glu Ser Ala Gln  
385 390 395 400

Arg Thr Leu Pro Val Val Leu Ala Arg Phe Arg Pro Leu Gly Val Leu  
405 410 415

Ala Arg Phe Arg Arg Leu Glu Gln Glu Thr Ala Gly Met Leu Leu Gly  
420 425 430

Asp Gln Glu Pro Glu Pro Arg Gly Phe Ile Ser Phe Thr Asp Phe Arg  
435 440 445

Asp Ser Asp Ala Phe Ala Ser Tyr Ala Glu Tyr Ala Ala Gln Phe Asn  
450 455 460

Asp Tyr Ile Asp Gln Tyr Ser Ile Leu Glu Ala Gln Arg Leu Ala Arg  
465 470 475 480

Ile Leu Ala Leu Gly Ser Arg Met Thr Val Asp Gln Trp Cys Leu Pro  
485 490 495

Leu Gln Lys Val Arg His Tyr Lys Val Leu Thr Ser Gln Pro Gly Leu  
500 505 510

Ile Ala Arg Gly Ile Glu Asn His Asn Arg Gly Ile Glu Tyr Cys Leu  
515 520 525

Gly Arg Pro Pro Leu Thr Asp Leu Pro Gly Leu Phe Thr Met Phe Gln  
530 535 540

Leu His Asp Ser Ser Trp Leu Leu Val Ser Asn Ile Asn Gly Glu Leu  
545 550 555 560

Trp Ser Asp Val Leu Ala Asn Ala Glu Val Met Gln Asn Pro Thr Leu  
565 570 575

Ala Ala Leu Ala Glu Pro Gln Gly Arg Phe Arg Thr Gly Arg Arg Thr  
580 585 590

Gly Gly Trp Phe Leu Gly Gly Pro Ala Thr Glu Gly Pro Ser Leu Arg  
595 600 605

Asp Asn Tyr Leu Leu Lys Leu Arg Gln Ser Asn Pro Gly Leu Asp Val  
610 615 620

Lys Lys Cys Trp Tyr Phe Gly Tyr Arg Gln Glu Tyr Arg Leu Pro Ala  
625 630 635 640

Gly Ala Leu Gly Val Pro Leu Phe Ala Val Ser Val Ala Leu Arg His  
645 650 655

Ser Leu Asp Asp Leu Ala Ala His Ala Lys Ser Ala Leu Tyr Lys Pro  
660 665 670

Ser Glu Trp Gln Lys Phe Ala Phe Trp Ile Val Pro Phe Tyr Arg Glu  
675 680 685

Ile Phe Phe Ser Thr Gln Asp Arg Ser Tyr Arg Val Asp Val Gly Ser  
690 695 700

Ile Val Phe Asp Ser Ile Ser Leu Leu Ala Ser Val Phe Ser Ile Gly  
705 710 715 720

Gly Lys Leu Gly Ser Phe Thr Arg Thr Gln Tyr Gly Asn Leu Arg Asn  
725 730 735

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Phe Val Val Arg Gln Arg Ile Ala Gly Leu Ser Gly Gln Arg Leu Trp  
                   740                  745                  750

Arg Ser Val Leu Lys Glu Leu Pro Ala Leu Ile Gly Ala Ser Gly Leu  
                   755                  760                  765

Arg Leu Ser Arg Ser Leu Leu Val Asp Leu Tyr Glu Ile Phe Glu Pro  
                   770                  775                  780

Val Pro Ile Arg Arg Leu Val Ala Gly Phe Val Ser Ala Thr Thr Val  
                   785                  790                  795                  800

Gly Gly Arg Asn Gln Ala Phe Leu Arg Gln Ala Phe Ser Ala Ala Ser  
                           805                  810                  815

Ser Ser Ala Gly Arg Thr Gly Gly Gln Leu Ala Ser Glu Trp Arg Met  
                   820                  825                  830

Ala Gly Val Asp Ala Thr Gly Leu Val Glu Ser Thr Ser Gly Gly Arg  
                   835                  840                  845

Phe Glu Gly Ile Tyr Thr Arg Gly Leu Gly Pro Leu Ser Glu Cys Thr  
                   850                  855                  860

Glu His Phe Ile Val Glu Ser Gly Asn Ala Tyr Arg Val Ile Trp Asp  
                   865                  870                  875                  880

Ala Tyr Thr His Gly Trp Arg Val Val Asn Gly Arg Leu Pro Pro Arg  
                           885                  890                  895

Leu Thr Tyr Thr Val Pro Val Arg Leu Asn Gly Gln Gly His Trp Glu  
                   900                  905                  910

Thr His Leu Asp Val Pro Gly Arg Gly Gly Ala Pro Glu Ile Phe Gly  
                   915                  920                  925

Arg Ile Arg Thr Arg Asn Leu Val Ala Leu Ala Ala Glu Gln Ala Ala  
                   930                  935                  940

Pro Met Arg Arg Leu Leu Asn Gln Ala Arg Arg Val Ala Leu Arg His  
                   945                  950                  955                  960

Ile Asp Thr Cys Arg Ser Arg Leu Ala Leu Pro Arg Ala Glu Ser Asp  
                           965                  970                  975

Met Asp Ala Ala Ile Arg Ile Phe Phe Gly Glu Pro Asp Ala Gly Leu  
                   980                  985                  990

Arg Gln Arg Ile Gly Arg Arg Leu Gln Glu Val Arg Ala Tyr Ile Gly  
                   995                  1000                  1005

Asp Leu Ser Pro Val Asn Asp Val Leu Tyr Arg Ala Gly Tyr Asp  
                   1010                  1015                  1020

Leu Asp Asp Val Ala Thr Leu Phe Asn Ala Val Asp Arg Asn Thr  
                   1025                  1030                  1035

Ser Leu Gly Arg Gln Ala Arg Met Glu Leu Tyr Leu Asp Ala Ile  
                   1040                  1045                  1050

Val Asp Leu His Ala Arg Leu Gly Tyr Glu Asn Ala Arg Phe Val  
                   1055                  1060                  1065

Asp Leu Met Ala Phe His Leu Leu Ser Leu Gly His Ala Ala Thr  
                   1070                  1075                  1080

Ala Ser Glu Val Val Glu Ala Val Ser Pro Arg Leu Leu Gly Asn  
                   1085                  1090                  1095

Val Phe Asp Ile Ser Asn Val Ala Gln Leu Glu Arg Gly Ile Gly  
                   1100                  1105                  1110

Asn Pro Ala Ser Thr Gly Leu Phe Val Met Leu Gly Ala Tyr Ser  
                   1115                  1120                  1125

Glu Ser Ser Pro Ala Ile Phe Gln Ser Phe Val Asn Asp Ile Phe  
                   1130                  1135                  1140



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Pro	Ala	Trp	Arg	Gln	Ala	Ser	Gly	Gly	Gly	Pro	Leu	Val	Trp	Asn
	1145					1150					1155			
Phe	Gly	Pro	Ala	Ala	Ile	Ser	Pro	Thr	Arg	Leu	Asp	Tyr	Ala	Asn
	1160					1165					1170			
Thr	Asp	Ile	Gly	Leu	Leu	Asn	His	Gly	Asp	Ile	Ser	Pro	Leu	Arg
	1175					1180					1185			
Ala	Arg	Pro	Pro	Leu	Gly	Gly	Arg	Arg	Asp	Ile	Asp	Leu	Pro	Pro
	1190					1195					1200			
Gly	Leu	Asp	Ile	Ser	Phe	Val	Arg	Tyr	Asp	Arg	Pro	Val	Arg	Met
	1205					1210					1215			
Ser	Ala	Pro	Arg	Ala	Leu	Asp	Ala	Ser	Val	Phe	Arg	Pro	Val	Asp
	1220					1225					1230			
Gly	Pro	Val	His	Gly	Tyr	Ile	Gln	Ser	Trp	Thr	Gly	Ala	Glu	Ile
	1235					1240					1245			
Glu	Tyr	Ala	Tyr	Gly	Ala	Pro	Ala	Ala	Ala	Arg	Glu	Val	Met	Leu
	1250					1255					1260			
Thr	Asp	Asn	Val	Arg	Ile	Ile	Ser	Ile	Glu	Asn	Gly	Asp	Glu	Gly
	1265					1270					1275			
Ala	Ile	Gly	Val	Arg	Val	Arg	Leu	Asp	Thr	Val	Pro	Val	Ala	Thr
	1280					1285					1290			
Pro	Leu	Ile	Leu	Thr	Gly	Gly	Ser	Leu	Ser	Gly	Cys	Thr	Thr	Met
	1295					1300					1305			
Val	Gly	Val	Lys	Glu	Gly	Tyr	Leu	Ala	Phe	Tyr	His	Thr	Gly	Lys
	1310					1315					1320			
Ser	Thr	Glu	Leu	Gly	Asp	Trp	Ala	Thr	Ala	Arg	Glu	Gly	Val	Gln
	1325					1330					1335			
Ala	Leu	Tyr	Gln	Ala	His	Leu	Ala	Met	Gly	Tyr	Ala	Pro	Ile	Ser
	1340					1345					1350			
Ile	Pro	Ala	Pro	Met	Arg	Asn	Asp	Asp	Leu	Val	Ser	Ile	Ala	Ala
	1355					1360					1365			
Thr	Tyr	Asp	Arg	Ala	Val	Ile	Ala	Tyr	Leu	Gly	Lys	Asp	Val	Pro
	1370					1375					1380			
Gly	Gly	Gly	Ser	Thr	Arg	Ile	Thr	Arg	His	Asp	Glu	Gly	Ala	Gly
	1385					1390					1395			
Ser	Val	Val	Ser	Phe	Asp	Tyr	Asn	Ala	Ala	Val	Gln	Ala	Ser	Ala
	1400					1405					1410			
Val	Pro	Arg	Leu	Gly	Gln	Val	Tyr	Val	Leu	Ile	Ser	Asn	Asp	Gly
	1415					1420					1425			
Gln	Gly	Ala	Arg	Ala	Val	Leu	Leu	Ala	Glu	Asp	Leu	Ala	Trp	Ala
	1430					1435					1440			
Gly	Ser	Gly	Ser	Ala	Leu	Asp	Val	Leu	Asn	Glu	Arg	Leu	Val	Thr
	1445					1450					1455			
Leu	Phe	Pro	Ala	Pro	Val									
	1460													

&lt;210&gt; SEQ ID NO 3

&lt;211&gt; LENGTH: 403

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Bordetella pertussis

&lt;400&gt; SEQUENCE: 3

Met	Ala	Pro	Leu	Leu	Val	Leu	Gly	Phe	Ala	Ser	Gly	Leu	Pro	Leu	Ala
1				5					10				15		

Leu	Ser	Ser	Gly	Thr	Leu	Gln	Ala	Trp	Ala	Thr	Val	Glu	Asn	Val	Ser
	20							25						30	

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Leu Gln Ser Ile Gly Phe Leu Thr Leu Ala Gly Thr Ala Tyr Thr Leu  
           35  40  45  
  
 Lys Phe Leu Trp Ala Pro Leu Ile Asp Arg Tyr Val Pro Pro Phe Leu  
           50  55  60  
  
 Gly Arg Arg Arg Gly Trp Met Leu Leu Thr Gln Val Leu Leu Ala Ala  
    65  70  75  80  
  
 Ala Ile Met Val Met Gly Met Leu Ser Pro Gly Ser Ala Leu Leu Pro  
   85  90  95  
  
 Leu Ala Leu Val Ala Val Leu Val Ala Phe Leu Ser Ala Ser Gln Asp  
   100  105  110  
  
 Ile Ala Phe Asp Ala Tyr Ser Thr Asp Val Leu Arg Gln Glu Glu Arg  
           115  120  125  
  
 Gly Ala Gly Ala Ala Met Arg Val Met Gly Tyr Arg Leu Ala Met Ile  
    130  135  140  
  
 Val Ser Gly Gly Leu Ala Leu Ile Val Ala Asp Arg Trp Leu Gly Trp  
    145  150  155  160  
  
 Gly Asn Thr Tyr Val Leu Met Gly Gly Leu Met Leu Ala Cys Ala Leu  
   165  170  175  
  
 Gly Thr Leu Trp Ala Pro Glu Pro Glu Arg Pro Ala Asn Pro Pro Arg  
   180  185  190  
  
 Asp Leu Gly Ala Ala Val Val Glu Pro Phe Arg Glu Phe Phe Ser Arg  
           195  200  205  
  
 Arg Gly Ala Ile Asp Met Leu Leu Leu Ile Val Leu Tyr Lys Leu Gly  
           210  215  220  
  
 Asp Ala Phe Ala Gly Ala Leu Ser Thr Thr Phe Leu Leu Arg Gly Ala  
    225  230  235  240  
  
 Gly Phe Ser Ala Thr Glu Val Gly Thr Val Asn Lys Val Leu Gly Leu  
   245  250  255  
  
 Ala Ala Thr Ile Val Gly Ala Leu Ala Gly Gly Ser Ile Met Thr Arg  
           260  265  270  
  
 Trp Gly Leu Tyr Arg Ser Leu Met Ala Phe Gly Leu Leu Gln Ala Val  
           275  280  285  
  
 Ser Asn Leu Gly Tyr Trp Leu Ile Ala Val Ser Pro Lys Asn Leu Tyr  
           290  295  300  
  
 Leu Met Gly Leu Ala Val Gly Val Glu Asn Leu Cys Gly Gly Leu Gly  
    305  310  315  320  
  
 Thr Ala Ser Phe Val Ala Leu Leu Met Ala Met Cys Arg Gln Gln Phe  
   325  330  335  
  
 Ser Ala Thr Gln Phe Ala Leu Leu Ser Ala Leu Ala Ala Val Gly Arg  
           340  345  350  
  
 Thr Tyr Leu Ala Gly Pro Leu Thr Pro Val Leu Val Glu Trp Leu Asp  
           355  360  365  
  
 Trp Pro Gly Phe Phe Ile Val Thr Val Leu Ile Ala Leu Pro Gly Leu  
           370  375  380  
  
 Trp Leu Leu Arg Leu Arg Arg Asn Val Ile Asp Glu Leu Asp Ala Gln  
    385  390  395  400  
  
 Thr Ala Arg

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 <211> LENGTH: 491  
 <212> TYPE: PRT  
 <213> ORGANISM: Escherichia coli

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&lt;400&gt; SEQUENCE: 4

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 20 25 30  
 Gly Thr Leu Gln Ala Trp Met Thr Val Glu Asn Ile Asp Leu Lys Thr  
 35 40 45  
 Ile Gly Phe Phe Ser Leu Val Gly Gln Ala Tyr Val Phe Lys Phe Leu  
 50 55 60  
 Trp Ser Pro Leu Met Asp Arg Tyr Thr Pro Pro Phe Phe Gly Arg Arg  
 65 70 75 80  
 Arg Gly Trp Leu Leu Ala Thr Gln Ile Leu Leu Leu Val Ala Ile Ala  
 85 90 95  
 Ala Met Gly Phe Leu Glu Pro Gly Thr Gln Leu Arg Trp Met Ala Ala  
 100 105 110  
 Leu Ala Val Val Ile Ala Phe Cys Ser Ala Ser Gln Asp Ile Val Phe  
 115 120 125  
 Asp Ala Trp Lys Thr Asp Val Leu Pro Ala Glu Glu Arg Gly Ala Gly  
 130 135 140  
 Ala Ala Ile Ser Val Leu Gly Tyr Arg Leu Gly Met Leu Val Ser Gly  
 145 150 155 160  
 Gly Leu Ala Leu Trp Leu Ala Asp Lys Trp Leu Gly Trp Gln Gly Met  
 165 170 175  
 Tyr Trp Leu Met Ala Ala Leu Leu Ile Pro Cys Ile Ile Ala Thr Leu  
 180 185 190  
 Leu Ala Pro Glu Pro Thr Asp Thr Ile Pro Val Pro Lys Thr Leu Glu  
 195 200 205  
 Gln Ala Val Val Ala Pro Leu Arg Asp Phe Phe Gly Arg Asn Asn Ala  
 210 215 220  
 Trp Leu Ile Leu Leu Leu Ile Val Leu Tyr Lys Leu Gly Asp Ala Phe  
 225 230 235 240  
 Ala Met Ser Leu Thr Thr Thr Phe Leu Ile Arg Gly Val Gly Phe Asp  
 245 250 255  
 Ala Gly Glu Val Gly Val Val Asn Lys Thr Leu Gly Leu Leu Ala Thr  
 260 265 270  
 Ile Val Gly Ala Leu Tyr Gly Gly Ile Leu Met Gln Arg Leu Ser Leu  
 275 280 285  
 Phe Arg Ala Leu Leu Ile Phe Gly Ile Leu Gln Gly Ala Ser Asn Ala  
 290 295 300  
 Gly Tyr Trp Leu Leu Ser Ile Thr Asp Lys His Leu Tyr Ser Met Gly  
 305 310 315 320  
 Ala Ala Val Phe Phe Glu Asn Leu Cys Gly Gly Met Gly Thr Ser Ala  
 325 330 335  
 Phe Val Ala Leu Leu Met Thr Leu Cys Asn Lys Ser Phe Ser Ala Thr  
 340 345 350  
 Gln Phe Ala Leu Leu Ser Ala Leu Ser Ala Val Gly Arg Val Tyr Val  
 355 360 365  
 Gly Pro Val Ala Gly Trp Phe Val Glu Ala His Gly Trp Ser Thr Phe  
 370 375 380  
 Tyr Leu Phe Ser Val Ala Ala Ala Val Pro Gly Leu Ile Leu Leu Leu  
 385 390 395 400

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Val Cys Arg Gln Thr Leu Glu Tyr Thr Arg Val Asn Asp Asn Phe Ile  
405 410 415

Ser Arg Thr Glu Tyr Pro Ala Gly Tyr Ala Phe Ala Met Trp Thr Leu  
420 425 430

Ala Ala Gly Ile Ser Leu Leu Ala Val Trp Leu Leu Leu Thr Met  
435 440 445

Asp Ala Leu Asp Leu Thr His Phe Ser Phe Leu Pro Ala Leu Leu Glu  
450 455 460

Val Gly Val Leu Val Ala Leu Ser Gly Val Val Leu Gly Gly Leu Leu  
465 470 475 480

Asp Tyr Leu Ala Leu Arg Lys Thr His Leu Met  
485 490

<210> SEQ ID NO 5  
<211> LENGTH: 32  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide A

<400> SEQUENCE: 5

tataaatcga tattcctgct ggtttcgctc tc 32

<210> SEQ ID NO 6  
<211> LENGTH: 30  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide B

<400> SEQUENCE: 6

tatagctagc aagttgggaa acgacaccac 30

<210> SEQ ID NO 7  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide C

<400> SEQUENCE: 7

taagaagcaa aataagccag gcatt 25

<210> SEQ ID NO 8  
<211> LENGTH: 30  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide D

<400> SEQUENCE: 8

tataccatgg cgccgctgct ggtgctgggc 30

<210> SEQ ID NO 9  
<211> LENGTH: 30  
<212> TYPE: DNA  
<213> ORGANISM: Artificial  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide E

<400> SEQUENCE: 9

tatatctaga cgctggccgt aaccttagca 30

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<210> SEQ ID NO 10
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Oligonucleotide F

<400> SEQUENCE: 10

tatagaattc gctcggttcg ctggtcaagg                30

<210> SEQ ID NO 11
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Oligonucleotide G

<400> SEQUENCE: 11

tatatctaga gcaatgccga ttcattctta                30

<210> SEQ ID NO 12
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Oligonucleotide H

<400> SEQUENCE: 12

tatatctaga gcggccttta ttgcttttcc                30

<210> SEQ ID NO 13
<211> LENGTH: 30
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Oligonucleotide I

<400> SEQUENCE: 13

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What is claimed is:

1. A method for protecting a subject against disease caused by *Bordetella* infection, comprising administering to the subject in need of such treatment a live attenuated *Bordetella pertussis* strain comprising at least a mutated *pertussis* toxin (ptx) gene, a deleted or mutated dermonecrotic (dnt) gene, and a heterologous ampG gene replacing the *Bordetella* ampG gene, wherein the strain is able to colonize and induce protective immunity in the subject.

2. The method of claim 1, wherein the *Bordetella* infection is a *Bordetella pertussis* infection.

3. The method of claim 1, wherein the subject is a newborn or a child.

4. The method of claim 1, wherein the live attenuated *Bordetella* strain is administered to the respiratory tract.

5. The method of claim 4, wherein the live attenuated *Bordetella* strain is administered intranasally.

6. The method of claim 1, wherein the live attenuated *Bordetella* strain is a BPZE1 strain deposited with the Collection Nationale de Culture Microorganismes as accession number 1-3585.

7. The method of claim 1, wherein the live attenuated *Bordetella* strain expresses less than 5% residual tracheal cytotoxin (CTC) activity.

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